

ANTIBIOGRAM ANALYSIS OF BOVINE MASTITIS-CAUSING BACTERIA ISOLATED FROM NEARBY FARMS OF LAHORE

Amina Rafique Shakir¹, Arslan Shaukat², Laiq Ahmed Athar Hussain Barvi³, Muhammad Jamal⁴, Anns Asgher⁵, Sudhair Abbas Bangash⁶, Hamna Sharif⁷, Ijaz Ahmad⁸, Safdar Aman⁹, Haseeb Khaliq¹⁰, Yafes Ali Shah^{11*}

¹Department of Microbiology and Molecular Genetics, Punjab University Lahore, Pakistan

²Department of Physiology, Government College University Faisalabad, Pakistan

³Veterinary Services Section, Public Health Services Department, Dubai Municipality, Dubai, United Arab Emirates

⁴Department of Biotechnology, University of Malakand, Khyber Pakhtunkhwa, Pakistan

⁵Department of Microbiology, University of Agriculture, Faisalabad, Pakistan

⁶Department of Pharmacy, Faculty of Life Sciences, Sarhad University of Science and Information Technology, Peshawar, Pakistan

⁷Department of Microbiology and Molecular Genetics, The Women University, Multan, Pakistan

⁸Department of Zoology, Khushal Khan Khattak University Karak, Khyber Pakhtunkhwa, Pakistan

⁹Department of Veterinary Medicine, University of Veterinary and Animal Sciences Lahore, Pakistan

¹⁰Department of Anatomy and Histology, Cholistan University of Veterinary and Animal Sciences, Bahawalpur, Pakistan

¹¹Department of Zoology, University of Science and Technology, Bannu, Pakistan
Email: yafesalishah@cug.edu.cn

Corresponding Author*: Department of Zoology, University of Science and Technology, Bannu, Pakistan, Email: yafesalishah@cug.edu.cn

Abstract:

Bovine mastitis has become major issue globally. This research is performed for investigating the current status of clinical mastitis among dairy cattle of farms of Lahore. The prevalence of mastitis was assessed by the results of the bacteriological evaluation of milk samples collected from clinical mastitis cases from farms in and nearby Lahore. Mastitic cattle and buffalos from farms nearby Lahore, Pakistan, have a high prevalence of multidrug-resistant bacteria. The main bacterial pathogen linked to mastitis was *Staphylococcus aureus*, followed by *E. coli* and *S. dysgalactiae*. This indicates that unhygienic and unsatisfactory management practices are being followed on the nearby farms. About 42 different mastitogenic bacteria including *Staphylococcus aureus* (28), *Escherichia coli* (12), and *Streptococcus dysgalactiae* (2) were isolated from the 47 confirmed clinical cases of mastitis affected cows and were further processed for antibiotic resistance profiling. The prevalence of major pathogens isolated was prevalence of 66.67%, 28.57%, and 4.76%, respectively. The results are similar to the earlier report of *S. aureus* (34%), *E. coli* (19%), and *Streptococci* spp. (9%) by Australia. Antibioqram analysis were also performed for these isolates shows that Gentamicin, Ciprofloxacin and Meropenem can be used as efficient antimicrobial agents for the treatment of bovine mastitis. This study has shown that microbiological and antibiogram analysis is very important for treatment and control of the mastitis.

Keywords: Bovine mastitis, AST, Antibioqram, microbiological analysis

Introduction:

Bovine mastitis has become a major issue globally. It is an inflammatory response of the udder tissue of cattle and buffalos resulted from some injury or microbial illness. It has become extremely

complex and the costliest disease in India. It affects 50% of the herd population¹. It has been estimated that the mastitis alone can cause approximately 70% of all avoidable losses incurred during milk production. It is believed to be the disease that causes the biggest financial damage to the dairy sector because of lower yield and poor milk quality. Bovine mastitis is thought to have a \$147 average annual cost per animal for failure overall. Approximately 11 to 18 percent of the gross margin per animal each year is contributed by milk production losses and 2. Injury to mammary tissue, which decreases milk production, is responsible for 70% of the total losses³.

Overuse of antibiotics without performing AST is the main reason for treatment failure⁽²⁾. This practice results in economic losses and development of AMR (antimicrobial resistance)⁽³⁾. The current study is intended to understand the antimicrobial resistance pattern (AMR) among the three most common mastitogenic bacteria including *S. aureus*, *Streptococcus* spp, and *E. coli* isolated from mastitic milk samples of infected bovines from various dairy farms of Lahore.

Material and Methods

1. **Collection of bacterial isolates:** For investigation, 60 milk samples were collected from clinical mastitis cases from different dairy farms located nearby Lahore. All information about the farm, breed and demographic history of each animal was recorded.
 2. **Media reagents and chemicals:** Different bacteriological grade media, such as mannitol salt agar, MacConkey agar, blood agar, and nutrient agar, were prepared in accordance with the manufacturer's instructions. The pH was adjusted using solutions of 10% NaOH and 10% HCL and were sterilized by autoclaving and then they were poured into glass Petri plates already sterilized by dry heat using hot air oven.
 3. **Identification of isolates:** *S. aureus*, *E. coli*, and *S. dysgalactiae* were streaked on mannitol salt agar, MacConkey agar, and blood agar, respectively. After being incubated at 37°C for 24-48 hours, the plates were assessed for growth as well as morphological features, including colony size, shape, color, and hemolytic traits. To obtain pure cultures, suspected colonies were chosen, and sub-cultured on nutrient agar. And then gram staining and biochemical tests (catalase and oxidase test) are performed for identification.
 4. **Antimicrobial susceptibility testing:** Minimal inhibition concentration (MIC) values of each isolated bacterium were analyzed. A panel of antimicrobials was prepared for each bacterial species. Amoxicillin-clavulanate potassium/ Augmentin (20/10µg), penicillin G (10units), gentamicin (10 µg), ciprofloxacin (5µg), clindamycin (2µg), chloramphenicol (30µg), and erythromycin (15µg) were selected for *Staphylococcus aureus*; augmentin (20/10µg), ampicillin (10µg), aztreonam (30µg), gentamicin (10µg), chloramphenicol (30µg), cefixime (5µg), ciprofloxacin (5µg), tetracycline (30µg), meropenem (10µg), and Co-trimoxazole (1.25/23.75µg), were used for *Escherichia coli*; and penicillin G (10units), vancomycin (30µg), erythromycin (15µg), tetracycline (30µg), chloramphenicol (30µg), ampicillin (10µg), linezolid (30µg), and clindamycin (2µg) were used for *S. dysgalactiae*. The zones of inhibition were then measured in mm and compared to CLSI standards for interpretation.
 5. **Broth Microdilution Method:**
The guidelines of the Clinical and Laboratory Standards Institute were followed for performing the broth microdilution method (Patel, Patel, Weinstein, Richter, & Eliopoulos, 2017).
- I. Preparation of Cation Adjusted Mueller Hinton Broth (CAMHB):**
21g of the dehydrated powder of Mueller Hinton broth (HIMEDIA) was dissolved in 1000mL of distilled water, pH was adjusted at 7.3±0.1 at 25° and sterilization by autoclaving. A tiny amount

of MHB medium was used to dissolve 20–25 mg of CaCl₂ and 10–12.5 mg of MgCl₂, separately. The solutions were then passed through a 0.2µm syringe filter and added to the medium. After the sterility test, it was used for the broth microdilution procedure.

II. Preparation of Stock Solution:

10mg of the lyophilized powder of vancomycin (Bosck Pharmaceuticals) was dissolved in 10mL of sterile distilled water and sterilized by passing through a 0.2µm syringe filter. It was then used for the preparation of working solution.

III. Preparation of Working Solution:

10mL of cation-adjusted Mueller Hinton broth (CAMHB) was taken in a sterile falcon tube and 25.6µL of the broth was replaced with the stock solution, according to the formula $C1V1=C2V2$. The resulting solution had 256µg of vancomycin in 10mL of CAMHB medium which was then used for making dilutions.

IV. Preparation of Bacterial Inoculum:

Three to four bacterial colonies from purified cultures were selected and transferred to 10mL of sterile normal saline in a test tube. 100µL from the suspension was transferred to a 96-wells microtiter plate and the OD was adjusted to 0.08-0.1 using an ELISA reader. Perform 10 fold dilution until the bacterial suspension became equivalent to 10⁵ CFU/mL.

V. Performance of Broth Microdilution Method:

200µL of the working solution was added to the first well of each row in a 96-wells microtiter plate. 100µL of cation-adjusted Mueller Hinton broth (CAMHB) was then added to the second well up till the last well of each row. 2-fold serial dilution was performed well till the 10th. Finally, 100µL of the bacterial suspension was added up to the eleventh well and zero day OD was measured at 630nm wavelength with the help of an ELISA reader. The 11th and 12th wells served as the respective positive and negative controls. After 24 hours of incubation at 37°C, the day one OD value was obtained, and ODnet was computed. The least concentration of vancomycin showing no obvious growth was taken as the minimum inhibitory concentration of the antibiotic. Finally, the results were interpreted as sensitive, resistant, or intermediate using the CLSI standards.

Results:

From 60 milk samples, 75 bacterial isolates were isolated. Out of 60 samples, pure cultures were isolated from 21 samples (35%). Out. OIILf which 6 were gram-positive and 15 were gram-negative organisms and the remaining 39 (65%) yielded mixed cultures. Of the 75 isolates, 49 (65.33%) were gram- positive and remaining 26 (34.67%) were gram-negative. The predominant bacterial isolates recovered were Staphylococcus aureus (24%) and Escherichia coli (20%) followed by Staphylococcus epidermidis(16%), Streptococcus sp. (16%), Klebsiella sp. (10.67%), Bacillus sp. (5.33%), Corynebacterium sp. (4%), Proteus sp. (2.66%) and Pseudomonas sp. (1.33%). The frequency of isolation of different bacterial species from clinical mastitis cases is depicted in Table1. The high prevalence of Staphylococcus species followed by E. coli in the present study is in accordance with work of several other workers.

Staphylococci are the most important and prevalent mastitis causing organism globally, including

India. Higher incidence of *E. coli* mastitis may be due to poor hygienic conditions, as *E. coli* originates from the cow's environment and infect the udder via the teat canal¹⁰. The in vitro antibiogram studies of the bacterial isolates from mastitis milk revealed gentamicin to be most effective drug (90%) followed by enrofloxacin (88%), ciprofloxacin (85%), chloramphenicol (75%), tetracycline (60%), colistin (57%), neomycin (50%), nitrofurantoin (50%), furazolidone (50%), cephalexin (47%), penicillin (45%), streptomycin (35%) and sulphadiazine (30%). Gentamicin, enrofloxacin, ciprofloxacin and chloramphenicol are newer chemotherapeutic agents and are less commonly used for treatment of mastitis in the area of study resulting in higher efficacy of these drugs. Gentamicin proved to be the drug of choice in this study. Similar antibiogram pattern were reported by other workers also^{12,13,14}. number of isolates showed resistance to cephalexin, penicillin, streptomycin and sulphadiazine. Indiscriminate and frequent use of these antibiotics in animals could be the reason for their ineffectiveness against bacterial isolates. Production of plasmids mediated beta-lactamase enzymes is supposed to be mainly responsible for resistance to penicillin. Since streptomycin has been extensively used along with penicillin for treating mastitis; it may have led to the development of high resistance in bacteria against this antibiotic. Whereas the resistance to sulphadiazine could be due to either low affinity of the enzyme that uses the p-amino benzoic acid during folic acid synthesis or use of preformed folic acid from surroundings.

Results and Discussion:

Different mastitogenic bacteria including *Staphylococcus aureus*, *Escherichia coli*, and *Streptococcus dysgalactiae* were isolated and cultured on various selective and differential media including mannitol salt agar, MacConkey agar, eosin methylene blue agar, and blood agar to obtain pure cultures. They were further confirmed through various phenotypic tests including Gram staining, catalase test, and oxidase tests to re-confirm the isolates. Antibiotic sensitivity assay was then performed through broth microdilution and disc diffusion assays to obtain an antibiogram.

Macroscopic Analysis:

Various selective and differential media were used for the macroscopic analysis of the isolates like mannitol salt agar was used for *Staphylococcus aureus*; MacConkey agar and eosin methylene blue agar were used for *Escherichia coli* and; blood agar was used for *Streptococcus dysgalactiae*. The colony morphology of the isolates is mentioned in table 4.1

Table 4.1: Colony Morphology of *S.aureus*, *E.coli*, and *S.dysgalactiae*

| Sr.No. | Bacterial Isolate | Medium | Size | Color | Shape |
|--------|-----------------------------------|---------------------------|-----------------|-------------------------|-------|
| 1. | <i>Staphylococcus aureus</i> | Mannitol Salt Agar | Small to Medium | Yellow | Round |
| 2. | <i>Streptococcus dysgalactiae</i> | Blood Agar | Pinpoint | White (Alpha Hemolytic) | Round |
| 3. | <i>Escherichia coli</i> | MacConkey Agar | Small | Pink | Round |
| 4. | <i>Escherichia coli</i> | Eosin Methylene Blue Agar | Medium | Green Metallic Sheen | Round |

Table 4.2: Microscopic characteristics of *S.aureus*, *E.coli*, and *S.dysgalactiae*

| Sr.No. | Bacterial Isolate | Gram staining | Shape | Arrangement |
|--------|-----------------------------------|---------------|---------|-------------|
| 1 | <i>Staphylococcus aureus</i> | G +ve | Cocci | Clusters |
| 2 | <i>Streptococcus dysgalactiae</i> | G +ve | Cocci | Chains |
| 3 | <i>Escherichia coli</i> | G -ve | Bacilli | Single |

Table 4.3: MDR Pattern of *S.aureus*, *E.coli*, and *S.dysgalactiae*

| Sr.No. | Bacteria | Number of Isolates | Resistance to Antibiotics N (%) | | | Multidrug Resistant Bacteria (%) |
|--------|-----------------------------------|--------------------|------------------------------------|---------------|----------------|----------------------------------|
| | | | 1 antibiotic | 2 antibiotics | >2 antibiotics | |
| 1 | <i>Staphylococcus aureus</i> | 28 | 3 (10.7%) | 12 (42.9%) | 13 (46.4%) | 89.3 |
| 2 | <i>Escherichia coli</i> | 12 | 0 (0%) | 0 (0%) | 12 (100%) | 100 |
| 3 | <i>Streptococcus dysgalactiae</i> | 2 | 0 (0%) | 0 (0%) | 2 (100%) | 100 |

| Sr.No. | Antibiotic | Concentration (µg) | Antibiotic Sensitivity Pattern | | |
|--------|-----------------|--------------------|--------------------------------|-------|-------|
| | | | S | I | R |
| 1 | Augmentin | 20/10 | 10.7% | 0% | 89.3% |
| 2 | Penicillin G | 10 units | 21.4% | 0% | 78.6% |
| 3 | Gentamicin | 10 | 92.9% | 0% | 7.1% |
| 4 | Ciprofloxacin | 5 | 89.3% | 10.7% | 0% |
| 5 | Clindamycin | 2 | 42.9% | 25% | 32.1% |
| 6 | Chloramphenicol | 30 | 71.4% | 17.9% | 10.7% |
| 7 | Erythromycin | 15 | 46.4% | 39.3% | 14.3% |

Table 4.4: Antibiotic sensitivity Pattern of *S.aureus*

Table 4.5: Antibiotic Susceptibility Pattern of *Streptococcus dysgalactiae*

| Sr.No. | Antibiotic | Concentration (µg) | Antibiotic Sensitivity Pattern | | |
|--------|-----------------|--------------------|--------------------------------|-------|-------|
| | | | S | I | R |
| 1 | Augmentin | 20/10 | 0% | 0% | 100% |
| 2 | Ampicillin | 10 | 0% | 0% | 100% |
| 3 | Aztreonam | 30 | 66.7% | 0% | 33.3% |
| 4 | Gentamicin | 10 | 66.6% | 16.7% | 16.7% |
| 5 | Chloramphenicol | 30 | 50% | 16.7% | 33.3% |
| 6 | Cefixime | 5 | 25% | 8.3% | 66.7% |
| 7 | Ciprofloxacin | 5 | 66.7% | 0% | 33.3% |
| 8 | Tetracycline | 30 | 0% | 0% | 100% |
| 9 | Meropenem | 10 | 91.7% | 8.3% | 0% |
| 10 | Co-trimoxazole | 1.25/23.75 | 50% | 8.3% | 41.7% |

Table 4.6: Antibiotic Susceptibility Pattern of *E.coli*

| Sr.No. | Antibiotic | Concentration (µg) | Antibiotic Sensitivity Pattern | | |
|--------|-----------------|--------------------|--------------------------------|-----|------|
| | | | S | I | R |
| 1 | Penicillin G | 10units | 50% | 0% | 50% |
| 2 | Vancomycin | 30 | 100% | 0% | 0% |
| 3 | Erythromycin | 15 | 100% | 0% | 0% |
| 4 | Tetracycline | 30 | 0% | 0% | 100% |
| 5 | Chloramphenicol | 30 | 50% | 50% | 0% |
| 6 | Ampicillin | 10 | 0% | 0% | 100% |
| 7 | Linezolid | 30 | 100% | 0% | 0% |
| 8 | Clindamycin | 2 | 50% | 0% | 50% |

In the current study, *S.aureus*, *E.coli*, and *S.dysgalactiae* were shown to be the main cause of bovine mastitis among the microbes obtained for the study with prevalence of 66.67%, 28.57%, and 4.76%, respectively. Since bacterial infection is the main cause of bovine mastitis, antimicrobial therapy is frequently used for its treatment. But antimicrobial resistance (AMR) has been

linked to low cure rates(7). According to the World Health Organization (WHO), antimicrobial resistance is accelerated by the misuse and overuse of antimicrobials(4).

In the study, over 78% of the *Staphylococcus aureus* were resistant to penicillin G (natural penicillin) while 21% of them were sensitive to it(14). Augmentin is also a β -lactam antibiotic that consists of amoxicillin along with clavulanate, a β -lactamase inhibitor. In the current study, over 89.28% of *S.aureus* were resistant to augmentin and 10.71% of them were sensitive to it. However, *E.coli* showed 100% resistance toward augmentin. Ampicillin, an aminopenicillin, is semi-synthetic northwest Pakistan that 86.1% of the mastitogenic bacteria had developed resistance to ampicillin (15). In this study, over 100 percent of *E.coli* and *S.dysgalactiae* were found resistant to ampicillin.

Aztreonam is a monobactam that interferes in the synthesis of bacterial cell wall by binding to an enzyme, called transpeptidase just like penicillin. In this study, about 66.7% of *E.coli* were found susceptible to aztreonam. Meropenem is a carbapenem and has the same mechanism of action as other β -lactam antibiotics. It has also shown good results against mastitogenic bacteria with a susceptibility rate of 91.7%. This demonstrates that aztreonam and meropenem can be really helpful in the treatment of bovine mastitis caused by *E.coli*.

Cefixime is a third-generation cephalosporin. Over 66.7% of *E.coli* were found resistant to cefixime antibiotic making it an ineffective way of managing bovine mastitis. Many other antibiotics like vancomycin are also used for the treatment of mastitis in bovines. In December 2016, Vancomycin Resistant *S.aureus* (VRSA) was reported in the milk of bovines and caprines. This was the first report of VRSA in food animals, despite the fact that the bacteria is frequently detected in humans. The detection of VRSA in milk is a huge concern because it could further threaten the health of its consumers.

In the research conducted in India, about two (0.73%) out of 274 isolates of *S.aureus* exhibited vancomycin resistance with MIC_{van} of 16 μ g/mL. While five isolates (1.83%) had shown intermediate response toward vancomycin with MIC_{van} of 8 μ g/mL(18).In this study, a total of ten *S.aureus* isolates were checked for vancomycin resistance. All (100%) had shown sensitivity toward vancomycin with MIC_{van} of 2 and 1 μ g/mL. There was not a single isolate that responded to vancomycin in an intermediate or resistance ranges. However, *S.dysgalactiae* was evaluated for vancomycin resistance and found 100% susceptible to it. In 2021, a study conducted in Pakistan showed that 77.7% of mastitogenic bacteria were resistant to erythromycin(15). While in this study, only 14.3% of *S.aureus* were found resistant to erythromycin. In the study, about 92.9% of *S.aureus*, and 66.6% of *E.coli* were susceptible to gentamicin. These results are similar to that of India (71.2%) (16), Poland (69.9%) (17),and northwest Pakistan (91.5%) (15). However, because of intrinsic resistance, aminoglycosides are not the preferred antimicrobials for the treatment of Streptococcal mastitis.

Tetracycline is a protein synthesis inhibitor. All the isolates of *E.coli* and *S.dysgalactiae* were found resistant to tetracycline in the study. Ciprofloxacin is a fluoroquinolon and a high proportion of *S.aureus* (89.3%), and *E.coli* isolates (66.7%) were found susceptible to ciprofloxacin.

Table 4.7: Mean Zone of Inhibitions and Minimum Inhibitory Concentration by *Staphylococcus aureus*, *Escherichia coli*, and *Streptococcus dysgalactiae*

| Sr.No. | Bacteria | Antibiotic | ZOI and MIC (Mean) |
|--------|-----------------------------------|-----------------|--------------------|
| 1 | <i>Staphylococcus aureus</i> | Penicillin G | 18.54 |
| | | Augmentin | 8.07 |
| | | Erythromycin | 22.11 |
| | | Clindamycin | 18.07 |
| | | Gentamicin | 24.46 |
| | | Ciprofloxacin | 26.5 |
| | | Chloramphenicol | 25.36 |
| | | Vancomycin | 15.20 |
| 2 | <i>Escherichia coli</i> | Ampicillin | 3.92 |
| | | Augmentin | 1.00 |
| | | Cefixime | 11.42 |
| | | Meropenem | 29.08 |
| | | Aztreonam | 23.75 |
| | | Gentamicin | 18.09 |
| | | Tetracycline | 2.17 |
| | | Ciprofloxacin | 23.00 |
| | | Co-trimoxazole | 14.83 |
| | | Chloramphenicol | 16.00 |
| 3 | <i>Streptococcus dysgalactiae</i> | Penicillin G | 22.5 |
| | | Ampicillin | 0.00 |
| | | Vancomycin | 19.5 |
| | | Erythromycin | 21.5 |
| | | Clindamycin | 9.5 |
| | | Tetracycline | 0.00 |
| | | Chloramphenicol | 21.5 |
| | | Linezolid | 26.5 |

Co-trimoxazole is a combination of sulphonamide and trimethoprim. In the study, about 12 isolates of *E.coli* were tested against co-trimoxazole. Five (41.7%) out of 12 isolates were resistant to the antibiotic while only one isolate (8.3%) gave an intermediate response to it (2).Chloramphenicol

binds to bacterial ribosomes, shows very good results so far with a susceptibility rate of 84.1% against mastitogens (15). Also, it was found that 71.4% of *S.aureus*, 50% of *E.coli*, and 50% of *S.dysgalactiae* were susceptible to chloramphenicol in the current study. Linezolid belongs to the class of oxazolidinones, the study showed 100% susceptibility for this antibiotic against *S.dysgalactiae*.

The MDR (Multidrug Resistance) trend mostly followed by the isolates include Augmentin-Ampicillin-Cefixime-Tetracycline-Ciprofloxacin-Cotrimoxazole-Chloramphenicol for *E.coli*; Penicillin G-Augmentin-Clindamycin-Vancomycin for *S.aureus*; and Ampicillin-Tetracycline for *S.dysgalactiae*. This trend is quite similar to the Penicillin-Augmentin-Vancomycin-Enrofloxacin-Tetracycline MDR trend for *S.aureus*; Augmentin-Tetracycline-Kanamycin-Cefquinome trend for *E.coli*; and Penicillin-Augmentin-Tetracycline trend for *Streptococcus* species reported by China (17).

The data was analyzed with the help of descriptive analysis by SPSS (version 20.0) and the results were compared with the standards of the Clinical and Laboratory Standards Institute (CLSI) and European Committee on Antimicrobial Susceptibility Testing (EUCAST), which provided the results for sensitive, intermediate, and resistant isolates, based on all mean values. According to the study, all the bacterial isolates were sensitive, intermediate, and resistant for the antibiotics. All the isolates were sensitive for gentamicin and ciprofloxacin while *E.coli* was also susceptible to the meropenem antibiotic. The results of the mean zone of inhibitions and mean minimum inhibitory concentration for the isolates is mentioned in table 4.7.

Mastitis therapy typically takes longer than expected because the illness recurs and the bacteria that cause it are difficult to eradicate even with a variety of antimicrobials. According to the World Health Organization a post-antibiotic era, far from being an apocalyptic fantasy, in which typical illnesses and minor wounds can kill is actually a very plausible possibility for the twenty-first century (16). So, in order to develop effective control measures and suitable therapeutic procedures, it is crucial to have knowledge of the prevalence of bovine mastitis and its causative agents.

Conclusion:

This study comes to the conclusion that mastitic cattle and buffalos from farms nearby Lahore, Pakistan, have a high prevalence of multidrug-resistant bacteria. The main bacterial pathogen linked to mastitis was *Staphylococcus aureus*, followed by *E.coli* and *S. dysgalactiae*. This indicates that unhygienic and unsatisfactory management practices are being followed on the nearby farms. Moreover, on the basis of the antimicrobial susceptibility profiling of the isolates, gentamicin, ciprofloxacin and meropenem can be used as efficient antimicrobial agents for the treatment of bovine mastitis.

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