

Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains

Hafiza Bisma Nadeem¹, Hafiz Khawar², Saira Jabeen², Maryam Salah ud Din², Mian Zahid sarfraz³, *Saba Abbas¹, Hira Shafique Awan⁴

¹Institute of Allied Health Sciences, School of Medical Laboratory Technology, Minhaj University Lahore,

²Dr. Ikram-ul-Haq Institute of Industrial Biotechnology, Government College University Lahore,

³ Department of Pathology, Allama Iqbal Medical College Lahore,

⁴ Department of Microbiology, University of Central Punjab Lahore

*Corresponding Author: Saba Abbas (sabaabbas786786@gmail.com)

ABSTRACT

Citrullus colocynthis belong to family Cucurbitaceae is a medicinal plant traditionally used for the treatment of various diseases [1][2] including diabetes, bacterial infections, constipation and many other diseases. Various parts of plant individually produced anti-microbial and anti-oxidant potential. This is the first study reporting 1. The Novel synergism effect of *Citrullus colocynthis*, Fruit, Seed and root extract against MDR pathogenic strains in the least concentration of various organic solvents extracted compounds (first study reporting anti-microbial activity of Butanol extracted compounds) 2. Inhibitory concentration (IC₅₀) of these compounds against MDR 3. Novel Synergism of *C. colocynthis* fruit, seed and root showing anti-oxidant potential. According to findings, *Citrullus colocynthis* can be used as medicinal plant as its various compounds isolated through organic solvents showed antimicrobial activity against MDR-pathogens. Maximum activity against MDR pathogens of *Pseudomonas Aeurignosa*, *Staphylococcus aureus* (MRSA), *Klebsiella pneumoniae*, *Stenotrophomonas maltophilia*, *Salmonella typhi*, *Salmonella paratyphi*, *Acinetobacter baumannii* and *Enterococcus faecalis* was shown by Butanol extract followed by Chloroform, Acetone, Ethanol, Hexane and DMSO extract. The plant also showed 92% anti-oxidant potential in the least concentration of 93µg/mL.

1.1 INTRODUCTION

Plants are named as medicinal plants due to presence of pharmacologically beneficial compounds and are being used worldwide from ancient time [1][2]. Almost 80% population worldwide use them as these plants exhibits medicinal properties due to presence of anti-oxidant [3] and antimicrobial properties [4][7] and many other properties due to secondary metabolites (alkaloids, glycosides, essential oils and terpenoids) [5] [6] which constitute the major portion of modern medicine by history of 50 years research [8] prepared from selection to grinding in consumable form under the supervision of experts[9], furthermore, these compounds are identified by GCMS and UPLC [10]

Citrullus colocynthis, member of Cucurbitaceae family has been traditionally used as medicinal plant from ancient time to aid the treatment of oedema [13], bacterial or fungal infection [14] constipation diabetes [20] and cancer [32]. The mature fruit possess 90% moisture with 30% protein, 10% carbohydrate 4% ash cover and 3% fiber content [14][36][37][39] ensuring the major portion of glycosides by the production of elataricin B, E and dihydroelataricin B [60][61].



Fig *Citrullus Colocynthis* Fruit (mature and immature)

Due to high temperature resistance property subtropical the plant is found in tropical and areas [59][22].

Regional language	Common Names
English	Colocynth or Vine-of-Sodom
Hindi	Indaryan or Ghorumba
Telugu	Eti-puchcha
Malayalam	Paikummati
Tamil	Paedikari- Attutummati
Kannada	Hamekkae, hamekkikayi
Arabic	Handhal
German	Bitter-melone or koloqnite
Spanish	Alhandhal or coloquintida
Sanskrit	Indarvani or Brihadvani
Bengali	Makhal, Indaryan, Panjot, Indrabuni
Gujrati	Indarayan
Marathi	Kaud-Indarvani
Punjabi	Kaudtumba
Urdu	Hanzal, Indaryan, Shahmehinzal
French	Coloquinte
Portuguese	Colocintida
Swedish	KoloKvint

The family shows genetic diversity due to extreme temperature resistance and efficiently grow in alkaline soil [55][56]. The water melon similar plant exhibit perennial structure due to herbaceous vine plant with yellow flower on axial site with apple sized extreme bitter fruit having white spongy flesh filled coriaceous peel with brown and white ovate seeds inside [57][58][59]

Hafiza Bisma Nadeem: Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains



Fig Morphological characteristics of *Citrullus colocynthis* fruit seed and pulp

KINGDOM	PLANTAE
Sub-kingdom	Tracheobionta
Super division	Spermatophyta
Division	Magnoliophyta
Class	Mangoliopsida
Sub class	Dillenidae
Order	Cucurbitales
Family	<i>Cucurbitaceae</i>
Genus	<i>Citrullus</i>
Specie	<i>Colocynthis (L.) Schard</i>

The plant shows pharmacological importance due to its secondary metabolites [15] with the treatment of various conditions including hyperlipidemia, analgesic ailment and diabetes [29][30][31]. The plant shows anti microbial [16][83][45], analgesic [24][32][28], anti cancerous [45][46][48], hypolipidimic [5][51], hypoglycemic [62][17][23][41][39][47][40], Larvicidal [52][53][54] anti alopecia [63][64] properties with the contrast of toxicity by due to its over consumption [25] leading severe condition [26] to various parts with possible cases of colitis, rectorrhagia and bloody diarrhea [34][35]. The plant shows its adverse affects due due to its anti fertility [64][65][66][67].

Resistance towards various antibiotics is a serious health care problem and is not only restricted to hospital environment as earlier due to over consumption of multi broad spectrum dosage [68][69] these include gram positive *S.aureus* and *Enterococcus* and gram negative ESBLs are being reported as major threat to develop antimicrobial resistance [69][70][71][72]. These MDR strains include *Pseudomonas Aeurignosa* MDR and XDR strains [73][74][75][76][77][78], *Stenotrophomonas maltophilia* [81][82][84], *Acinetobacter baumanii* [79][80], *Staphylococcus aureus* (MRSA) [85][86][87][89], *Salmonella typhi* [90][91][92][93][94], *Salmonella paratyphi* [95][96][97], *Klebsiella pneumoniae* with it MDR-hvKP strains [99][100][101] with poor MDR strains of *Enterococcus faecalis* [102][103][104][105]

The study was designed to evaluate whether *Citrullus colocynthis* can inhibit the growth of MDR pathogenic strains as reported in earlier studies which include its antimicrobial property against ATCC strains and other lab strains as well as evaluation of their least effective dose working as IC₅₀ against them using the Novel synergism of its own fruit seed and root which individually shown the anti microbial property.

Materials

Bacterial MDR clinical isolates, Sterile swabs, Petri dishes, Wire loop, Bunsen burner, Eppendorf tubes, Glass tubes, Surgical gauze, Micro pipette 100-200 μ L, Micro pipette 500-100 μ L, Sterile tips, Glass tubes, Beaker (50, 100, 250, 500 and 100)ml, Conical flask (500 and 1000ml), Spatula, Separating funnel, Watman filter paper no.41, Aluminium Foil, Water bath, Safety cabinet, Micro titer plates, Incubator, Shaking incubator, Weighing Balance (0.000g) , Marker (permanent), Match box, Fridge for storage, Sanitizer, Cotton, Sterile Gloves, Measuring Cylinder, Paraffin tape, Wooden sticks, MacConkey agar, Mannitol salt agar, Mueller Hinton agar, Nutrient agar, Glycerol, Distilled water, Trypton, NaCl, Beef extract, Yeast extract, Normal Saline (0.5% NaCl), 70% ethanol, Cefexime, DPPH, DMSO, n-Hexane, Butanol, Methanol, Ethanol, Acetone, Ethyl Acetate, Cat-ion adjusted MH Broth

Methodology

Plant Preparation

Plant material was collected from tropical region of Pakistan and the sample was identified by the head of botany department Minhaj University Lahore. After identification the plant was sterilized after washing with deionized water and allow drying in hot air oven for 3-7 days within temperature of 35-37°C.

Plant Extraction

After drying the plant extract was grinded into fine powder and was measured using weighing balance was mixed with 1000ml methanol allowing extraction at 25°C for 15-20 days

Crude preparation

Using a surgical gauze the extract was filter out as pure solution of plant metabolic compounds in methanol using a surgical gauze and watman filter paper no. 41 and allowing crude preparation by standing on water bath at 60°C to get semisolid crude extract

Fractionation

Fractionation was done using various organic solvents by 2 solvent fractionation by using 1:2 by using Chloroform. Ethanol, DMSO, Butanol, n-Hexane and Acetone. After extraction in various organic solvents all the solvents were allowed to dry on water bath by keeping the temperature range below than their boiling point

Evaluation of Antimicrobial Potential

Collection of Bacterial Isolates

MDR strains were collected from Sheikh Zayed Hospital Lahore under the supervision of Head of microbiology Department Sheikh Zayed Hospital Lahore the MDR strains of *Staphylococcus aureus* (MRSA), *Klebsiella Pneumoniae*, *Acinetobacter Bummaniae*, *Salmonella Typhi*, *Salmonella Paratyphi*, *Enterococcus faecalis*, *Pseudomonas aeruginosa* and *Stenotrophomonas Maltophilia* were collected using sterile swabs

Hafiza Bisma Nadeem: Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains

Isolation of Pure colonies

pure colonies of these isolates were obtained using manitol salt agar and macConkey agar plates for gram positive and negative respectively. The safety cabinet was sterilized with 70% ethanol and leave under UVLight for 15 minutes after and prior the work and place the

Evaluation of antibiogram

For the confirmation MDR strain, anti-biogram of these MDR strains was evaluated by using Disc Diffusion method and their resistance pattern toward various antibiotics was documented

Table : Antibiogram data of MDR pathogens collected from Diagnostic Cultures at Shiekh Zayed Hospital Lahore

antibiotic	p.aeurignosa	S.aureus	k.pneumoniae	S.Maltophilia	S.typhi	S.paratyphi	A.baumani	E.faecalis
AK	R	R	-	R	-	-	R	I
AMC	-	-	R	-	-	-	-	-
AMP	-	-	S	-	R	R	R	-
ATM	S	-	R	R	-	-	-	-
AZM	-	-	-	-	S	S	-	-
C	-	I	S	-	R	S	R	R
CAZ	R	-	R	R	-	-	R	-
CIP	R	I	R	S	R	R	S	S
CN	R	R	S	S	-	-	I	S
CRO	-	-	R	-	-	-	-	-
CXM	-	-	S	-	-	-	S	-
DA	-	R	-	-	-	-	-	S
E	-	S	R	-	-	-	R	S
FD	-	R	R	-	-	-	-	R
FOT	-	-	R	-	-	-	-	-
FOX	-	R	-	R	-	-	-	S
IPM	R	-	R	S	-	-	R	-
LEV	-	-	-	S	-	-	-	-
MEM	R	-	S	S	S	S	S	-
P	-	R	-	-	-	-	-	R
PB	S	-	-	R	-	-	R	-
SXT	R	R	S	R	-	-	R	S
TE	-	S	R	-	-	-	-	S
TEG	-	-	R	-	-	-	R	-
TOB	R	-	-	S	-	-	I	-
TZP	R	-	S	S	-	-	-	-
VA	-	S	-	-	-	-	-	R

Preparation of Culture Broth

Culture broth was prepared for the purpose of storage as well as for the purpose of MIC work. Freshly prepared nutrient broth was used

Storage of Bacteria

Bacteria were stored as they were collected in different periods using 50% glycerol solution freshly prepared in lab. Using sterile eppendorf tubes all the organisms were stored in their glycerol stock and stored at -80°C for the future use.

Preparation of Antibiotic Stock Solution

To Determine the antimicrobial activity and MIC it was necessary to obtain liquid drugs antibiotic stock solution were prepared by adding 3mg of semisolid extract in their respective solvents producing 3mg/ml antibiotic stock solution of each antibiotic solvents of ethanol, Butanol, chloroform, acetone, n-Hexane and DMSO. All the solution were kept in fridge and taken out at least 30 minutes prior to application

Preparation of Nutrient Agar

Nutrient agar was only prepared to obtain the fresh growth from freeze stock microbes stock. These organism were streak on nutrient agar using sterile wooden sticks

Setting Up MIC apparatus

MIC set up was done by using Micro titer plate labeled with the applied antibiotic stock on the top of all rows the concentration was mentioned in the range between 300 to $1.17\mu\text{g/ml}$ using 2 fold serial dilution while the first column serve as antibiotic control (Cephalosporin 200mg/ml). and the 2nd last column served as growth control while the last column served as sterility control. Cation adjusted Mueller Hinton agar was used as media. Using the MIC method each well was served with $100\mu\text{L}$ of Mueller hinton broth. Wells of 1st row were served with antibiotic control while the other 9 columns showed the descending concentration of test solvents. The growth control column only consist of bacterial isolate and Mueller hinton broth confirming the test free from false positive results while the sterility column contain only broth confirming test free from environmental contamination.

After applying all the setup each plate was covered with sterile lid and incubated at 37°C with 5% CO_2 for 24 hours.

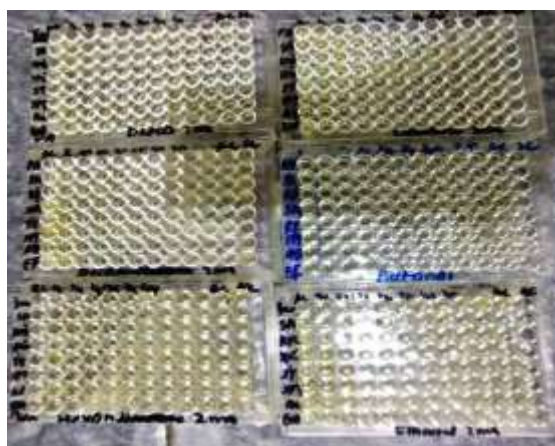


Fig Micro dilution Method of Evaluation of MIC of MDR Strains

After the completion of incubation period the results were determined using ELISA plate reader. Triplicates of reading were taken and data was analyzed to determine the percentage inhibition by using the formula of

$$\frac{\text{Absorbance of control (Ac)} - \text{Absorbance of sample (As)}}{\text{Absorbance of control (Ac)}} \times 100$$

Hafiza Bisma Nadeem: Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains

These percentage were further analyzed in comparison with one another

Preparation of Crude for Anti-oxidant potential determination

For this purpose the methanolic extract was diluted into 5 different concentrations of 1500, 750, 375, 187.5 and 93.75 by two fold serial dilution method in order to check the radical scavenging potential against DPPH free radicals determining its inhibitory potential ratio at various concentration

DPPH Preparation

0.1mM DPPH was prepared by dissolving 2mg DPPH in 250ml methanol which was further homogenized in ultrasonic bath for approximately 30 sec and was stored in dark (Aluminum Foil wrapped flask) due to its light sensitivity

DPPH Assay

Using an ELISA plate DPPH assay was tested to evaluate anti-oxidant potential of Synergism of *C.colocynthis* each well was marked with the methanolic concentration used in it i.e. 1500, 750, 375, 187.5 and 93.75 $\mu\text{L}/\text{mL}$ present in the volume of 100 μL . after incubation with 0.1mM DPPH in the period of 30 minutes results was evaluated with ELISA plate reader of BioTEK 800 TS. And results were documented in the percentage inhibition using the standard formula after taking triplicate of reading.

$$\frac{\text{Absorbance of control (Ac)} - \text{Absorbance of sample (As)}}{\text{Absorbance of control (AC)}} \times 100$$

RESULTS

Concentration ($\mu\text{g}/\text{mL}$)	Inhibition (%)
1500	93.87 \pm 0.25
750	93.50 \pm 0.93
375	93.38 \pm 0.58
187.5	92.84 \pm 0.46
93.75	92.76 \pm 0.35

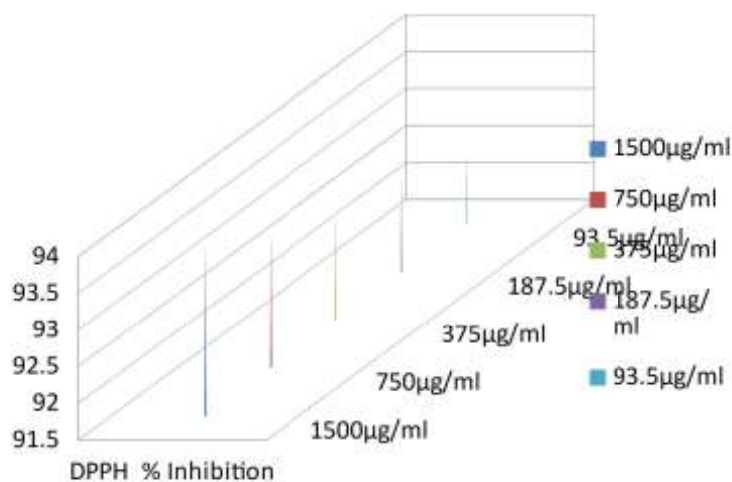


Fig Graphical representation of Maximum %inhibition of Methanol extract of C.colocynthis against
DPPH

Table : Mean percentage inhibition of Citrullus Colocynthis Ethanol extract Against MDR Pathogens

Mean% Inhibition Of MDR Pathogens								
Conc. µg/ml	<i>P.aeruginosa</i>	<i>S.aureus</i>	<i>K.pneumoniae</i>	<i>S.maltophilia</i>	<i>S. typhi</i>	<i>S.paratyphi</i>	<i>A.baumannii</i>	<i>E.faecalis</i>
300	56.82±0.54	62.31±0.15	75.73±0.41	60.83±0.55	59.86±1.09	52.97±0.62	75±0.08	59.43±1.01
150	55.08±0.35	56.92±0.42	73.34±0.64	57.14±1.50	54.71±0.87	51.14±1.67	71.06±1.69	56.44±1.57
75	52.55±1.28	55.41±1.92	72.42±0.69	53.41±1.52	51.85±1.36	48.47±0.31	68.34±0.93	51.29±1.54
37.5	50.16±0.36	53.38±0.74	70.46±0.32	50.55±0.99	44.89±1.03	46.95±0.83	63.44±0.71	44.61±1.90
18.75	49.74±0.66	50.25±0.42	67.83±0.57	45.80±0.86	41.26±1.18	43.83±0.73	57.04±1.50	39.42±1.53
9.37	47.10±0.98	47.02±0.62	63.90±0.78	42.22±0.62	35.26±0.99	39.95±0.93	48.39±2.21	30.26±0.41
4.68	42.10±0.12	43.85±0.53	59.65±1.99	40.20±0.50	32.05±0.26	38.91±0.36	42.02±1.89	28.85±1.30
2.34	41.20±2.70	39.59±1.2	54.78±1.2	35.80±1.2	29.31±1.2	33.50±1.2	30.43±1.2	21.93±1.2
1.2	35.20±1.2	25.22±0.22	48.33±0.78	30.45±1.24	20.16±1.56	28.33±1.23	21.25±1.23	16.51±0.85

Mean percentage inhibition of Citrullus Colocynthis Hexane extract A

Hafiza Bisma Nadeem: Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains

Mean% Inhibition Of MDR Pathogens								
Conc. µg/ml	<i>P.aeruginosa</i>	<i>S.aureus</i>	<i>K.pneumoniae</i>	<i>S.maltophilia</i>	<i>S. typhi</i>	<i>S.paratyphi</i>	<i>A.baumannii</i>	<i>E.faecalis</i>
300	49.88±0.34	60.16±1.67	45.74±0.1	42.08±0.3	40.52±1.0	54.53±1.2	64.0±1.0	50.0±1.5
150	48.42±0.54	58.99±1.31	42.18±0.42	38.02±0.97	33.13±2.9	49.48±1.3	59.01±2.2	44.41±1.7
75	45.05±0.54	55.35±1.01	37.5±1.7	32.68±1.0	30.62±2.25	46.63±1.1	56.83±0.5	40.17±1.14
37.5	41.73±0.69	53.88±0.82	34.98±0.66	31.70±0.36	25.44±0.7	40.88±2.2	52.18±1.2	31.32±1.0
18.75	40.38±0.38	51.94±1.13	32.04±1.5	28.25±0.65	19.76±0.81	38.53±0.9	48.42±0.8	28.10±1.14
9.37	37.21±0.31	46.58±1.38	27.91±1.07	21.38±0.86	18.71±1.0	33.74±1.19	44.09±1.86	24.01±1.01
4.68	34.26±1.35	44.92± 1.5	26.07±0.46	16.50±1.6	13.22±0.6	27.40±0.3	38.48±2.6	18.55±0.74
2.34	32.82±0.01	40.10±1.2	24.11±0.8	15.29±0.6	7.23±102	23.38±1.2	30.68±1.2	17.01±0.42

Table Mean percentage inhibition of *Citrullus Colocynthis* DMSO extract Against MDR Pathogens

Mean% Inhibition Of MDR Pathogens								
Conc. µg/ml	<i>P.aeruginosa</i>	<i>S.aureus</i>	<i>K.pneumoniae</i>	<i>S.maltophilia</i>	<i>S. typhi</i>	<i>S.paratyphi</i>	<i>A.baumannii</i>	<i>E.faecalis</i>
300	52.50±0.4	61.68±0.53	34.55±0.72	50.19±0.17	52.24±0.55	36.41±0.66	56.08±0.56	54.44±0.44
150	50.19±0.34	57.42±0.40	31.98±0.45	41.99±0.44	45.78±1.27	22.38±1.86	46.63±0.22	51.10±0.0.29
75	44.35±0.55	50.25±2.99	27.05±0.47	36.88±1.13	40.44±0.17	17.46±0.57	43.52±1.01	48.03±0.60
37.5	39.85±0.9	40.47±2.38	25.82±0.66	30.07±1.12	35.42±0.70	16.11±0.36	30.40±3.51	42.76±0.64
18.75	38.11±0.46	36.05±0.69	22.08±0.23	26.0±0.24	30.39±0.58	11.64±0.51	29.97±0.73	40.48±0.75
9.37	36.34±0.42	34.39±0.28	18.22±1.18	20.67±0.62	19.56±0.65	8.24±1.63	24.92±0.73	33.88±1.95
4.68	34.02±0.21	26.19±0.05	14.98±0.93	12.56±1.68	13.30±3.19	6.13±0.20	17.56±1.82	27.63±1.65
2.34	30.80±1.23	18.48±1.03	4.74±0.42	1.46±1.08	2.47±2.01	1.24±0.57	11.76±0.75	17.21±0.35
1.2	24.44±1.35	12.36±1.15	3.45±0.58	resistant	resistant	resistant	2.74±1.25	10.23±0.74

Table: Mean percentage inhibition of *Citrullus Colocynthis* Chloroform extract Against MDR Pathogen

Mean% Inhibition Of MDR Pathogens								
Conc. µg/ml	<i>P.aeruginosa</i>	<i>S.aureus</i>	<i>K.pneumoniae</i>	<i>S.maltophilia</i>	<i>S. typhi</i>	<i>S.paratyphi</i>	<i>A.baumannii</i>	<i>E.faecalis</i>
300	69.75±1.79	54.38±0.29	64.0±1.08	65.20±0.24	61.29±1.81	64.91±0.62	71.99±0.38	59.39±2.19
150	66.63±0.35	53.97±1.68	60.41±2.11	61.03±0.19	54.17±0.61	59.39±1.14	68.24±0.53	52.90±0.18
75	64.89±1.01	50.40±2.54	54.81±0.59	57.09±1.20	50.41±1.05	56.47±0.42	63.69±2.23	50.0±0.54
37.5	59.69±0.75	45.39±1.29	52.69±1.04	51.69±0.99	42.42±0.74	51.31±1.04	58.47±1.08	46.14±1.42
18.75	57.50±0.52	41.51±1.76	47.64±1.45	48.04±1.28	39.94±1.79	49.61±2.58	56.18±1.55	42.45±1.24
9.37	54.15±1.65	40.57±1.27	44.76±0.15	42.38±0.77	33.72±3.76	46.15±0.31	49.21±1.79	39.11±1.55
4.68	50.05±0.25	35.58±1.22	40.59±1.10	39.87±1.04	26.37±3.46	41.82±1.74	46.17±1.77	36.04±1.75
2.34	42.74±0.65	30.88±0.25	36.12±1.12	35.12±0.12	19.05±1.87	38.0±0.02	40.37±1.35	30.71±1.04

Table : Mean percentage inhibition of Citrullus Colocynthis Acetone extract Against MDR Pathogens

Mean% Inhibition Of MDR Pathogens								
Conc. µg/ml	<i>P.aeruginosa</i>	<i>S.aureus</i>	<i>K.pneumoniae</i>	<i>S.maltophilia</i>	<i>S. typhi</i>	<i>S.paratyphi</i>	<i>A.baumannii</i>	<i>E.faecalis</i>
300	57.24±0.10	60.40±1.55	51.07±0.28	65.82±1.80	51.35±1.04	55.61±2.04	58.86±0.56	53.10±0.33
150	56.21±0.43	57.70±0.58	46.13±0.97	61.52±0.33	43.07±0.4	52.14±0.33	57.43±0.55	48.89±2.61
75	53.09±0.92	55.45±1.13	42.49±0.78	57.29±0.84	39.59±0.87	48.40±1.78	49.78±1.50	41.86±0.82
37.5	50.25±0.36	52.94±0.35	37.28±0.34	53.02±0.70	34.22±1.33	42.65±1.47	43.95±1.03	36.94±1.44
18.75	48.87±0.39	48.87±0.52	34.46±0.95	49.80±1.12	31.01±0.82	38.11±0.42	36.98±0.48	28.30±1.39
9.37	44.15±1.06	38.15±1.98	27.78±1.13	42.77±1.03	24.12±0.99	33.71±2.42	35.58±0.43	19.10±1.0
4.68	40.97±0.08	26.0±0.63	24.54±1.03	38.93±1.20	18.75±0.46	27.26±0.83	32.40±0.70	15.72±0.29
2.34	33.95±0.50	20.73±1.05	18.13±0.34	33.0±0.99	11.05±1.74	21.41±1.98	28.21±2.57	6.80±0.59
1.2	19.98±1.42	8.33±2.75	6.04±1.58	21.25±0.33	5.36±0.23	12.82±0.45	16.86±1.78	resistant

Table: Mean percentage inhibition of Citrullus Colocynthis Butanol extract Against MDR Pathogens

Hafiza Bisma Nadeem: Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains

Mean% Inhibition Of MDR Pathogens

Conc. µg/ml	<i>P.aeruginosa</i>	<i>S.aureus</i>	<i>K.pneumoniae</i>	<i>S.maltophilia</i>	<i>S. typhi</i>	<i>S.paratyphi</i>	<i>A.baumannii</i>	<i>E.faecalis</i>
300	92.46±1.06	92.10±0.93	92.70±0.377	90.75±0.75	89.44±0.71	91.71±0.42	91.55±0.528	91.11±0.24
150	88.33±0.45	81.89±0.56	86.82±0.47	86.62±0.51	85.61±2.79	84.61±0.64	82.72±0.67	84.59±2.7
75	86.93±0.46	79.38±1.41	83.85±.47	79.45±1.66	82.63±0.48	79.97±.26	80.07±1.22	76.49±0.18
37.5	83.50±0.42	77.13±.81	82.23±1.06	75.97±1.52	79.69±1.53	77.68±1.19	80.90±.53	70.95±1.30
18.75	81.15±1.01	68.79±1.38	78.79±1.24	71.19±0.67	75.32±1.40	72.79±1.02	77.25±0.70	65.60±0.44
9.37	70.26±1.11	63.69±1.41	73.28±0.70	65.13±1.12	68.94±1.28	70.16±0.74	72.42±1.23	63.01±1.42
4.68	66.30±0.17	60.15±.80	68.36±0.93	60.97±0.20	65.54±0.61	67.15±0.68	69.31±1.39	57.94±1.41
2.34	57.39±0.71	51.78±0.95	63.05±0.59	57.94±2.65	62.37±1.24	63.99±2.02	66.66±2.12	55.30±0.04
1.2	51.23±1.02	41.23±1.23	54.26±0.48	49.23±2.56	54.01±0.56	55.87±1.58	57.82±2.87	48.56±0.63

Fig.4.2: Graphical %inhibition of all against Pseudomonas

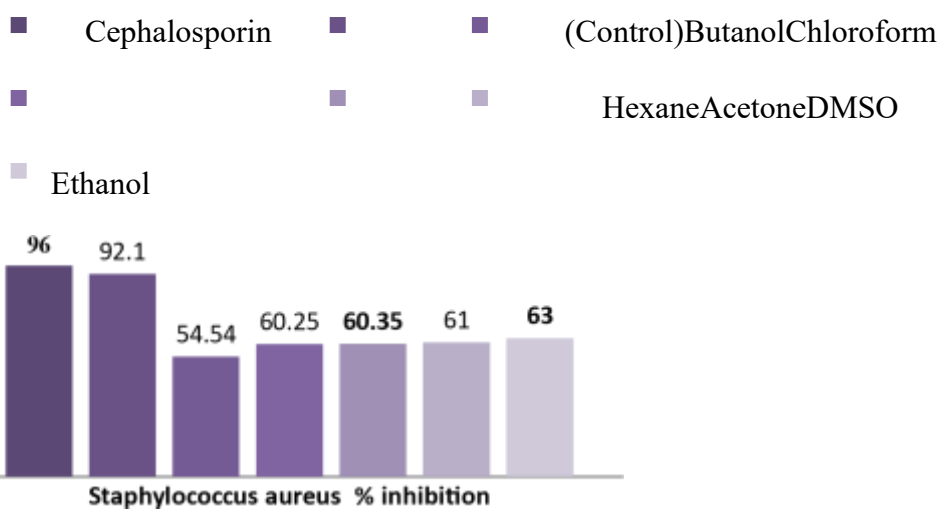
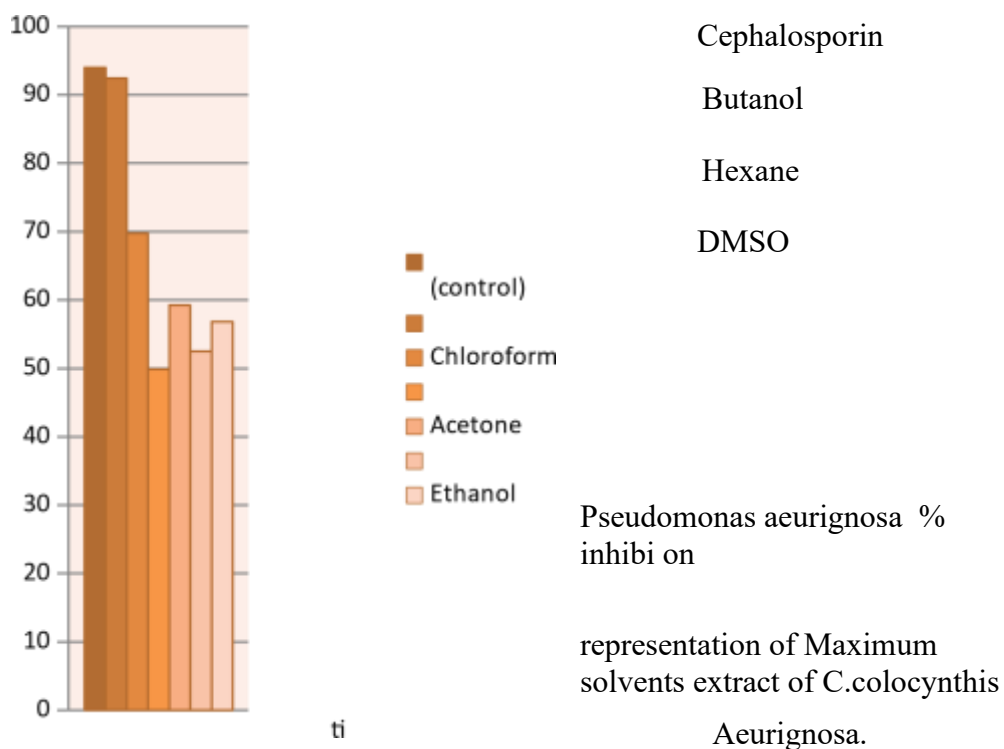


Fig.4.3: Graphical representation of Maximum %inhibition of all solvents extract of C.colocynthis against Staphylococcus Aureus (MRSA).

Hafiza Bisma Nadeem: Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains

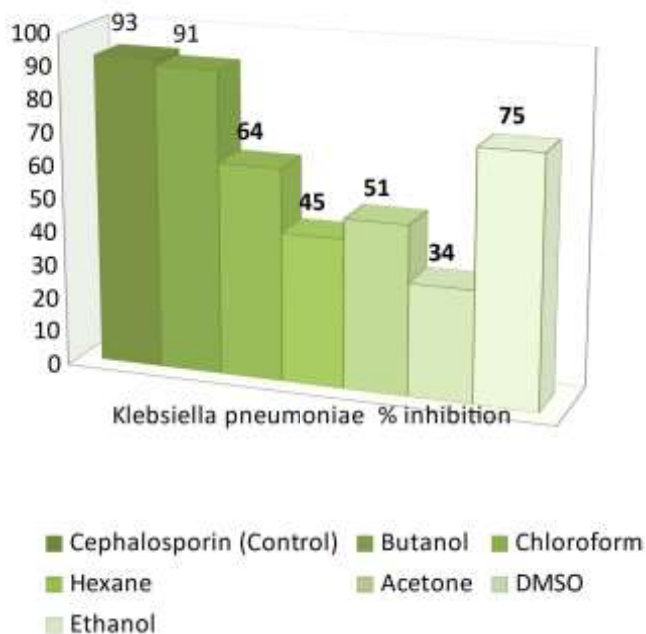


Fig.4.3: Graphical representation of Maximum %inhibition of all solvents extract of *C.colocynthis* against *Klebsiella pneumoniae*.

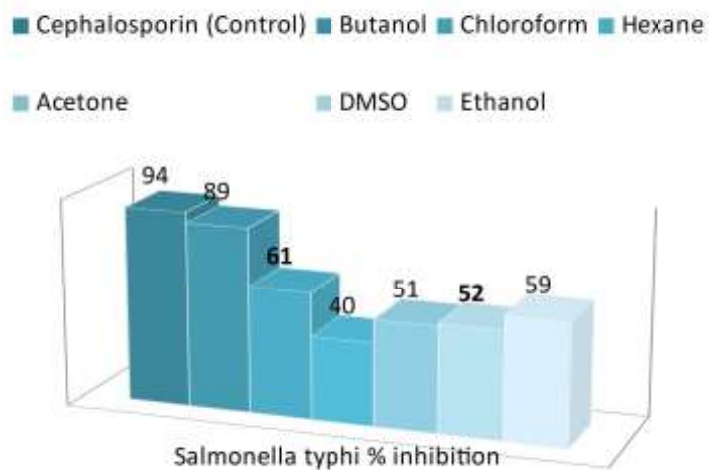


Fig.4.4: Graphical representation of Maximum %inhibition of all solvents extract of *C.colocynthis* against *Salmonella Typhi*.

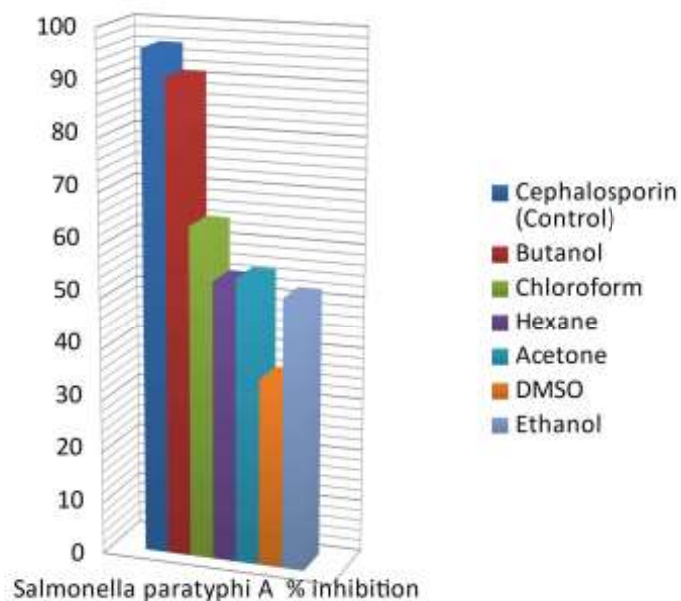


Fig.4.5: Graphical representation of Maximum %inhibition of all solvents extract of C.colocynthis against Salmonella Paratyphi A.

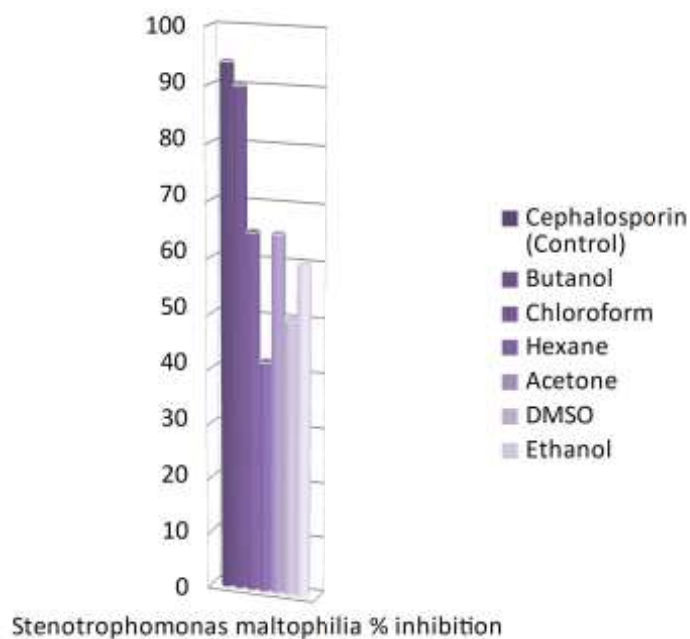


Fig.4.6: Graphical representation of Maximum %inhibition of all solvents extract of C.colocynthis against Stenotrophomonas maltophilia.

Hafiza Bisma Nadeem: Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains

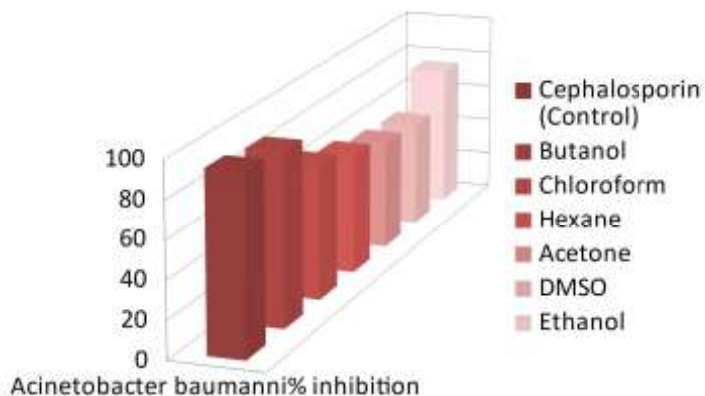


Fig.4.7: Graphical representation of Maximum %inhibition of all solvents extract of *C.colocynthis* against *Acinetobacter baumannii*.

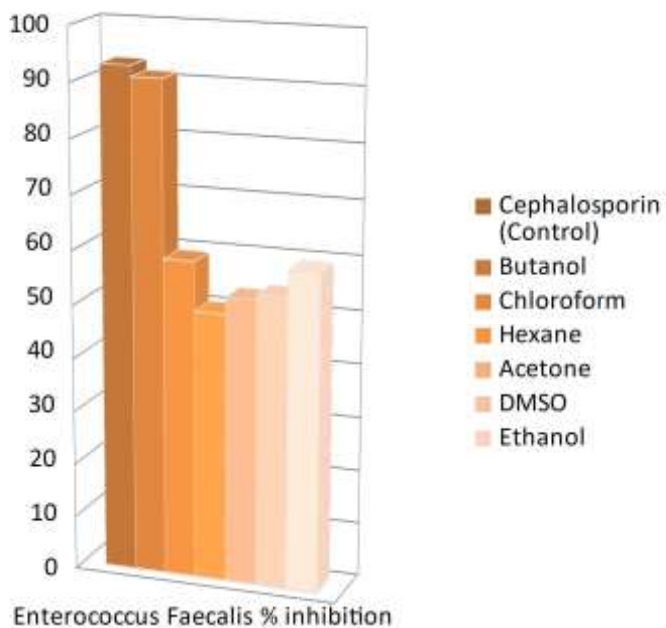


Fig.4.2: Graphical representation of Maximum %inhibition of all solvents extract of *C.colocynthis* against *Enterococcus Faecalis*.

Table 4.8: Mean growth inhibition (%) of MDR strains by Control (third generation Cephalosporin)

MDR Strains	Inhibition (%)
Pseudomonas Aeurignosa	94±0.35
Staphylococcus aureus(MRSA)	96±0.96
Klebsiella Pneumoniae	93±0.74
Stenotrophomonas Maltophilia	94±1.2
Salmonella Typhi	91±1.32
Salmonella Paratyphi A	96±0.03
Acinetobacter baumannii	94±0.26
Enterococcus Faecalis	93±0.99

Discussion

The study proves the enhanced or effective result of synergism effect of *Citrullus colocynthis* root, seed and fruit in both ways including anti-oxidant and anti microbial activity as they show both phenomenon. The goal of the study was to evaluate the least effective concentration used to inhibit the growth by keeping in mind the Plant's lethal dose. For anti-microbial activity, The novelty of the work is that the compounds isolated in Butanol organic solvent showed the maximum inhibitory affect of 91% against MDR strains in the least concentration of 1.2µg/mL ensuring its pharmaceutical importance. Not all the solvent extracted compound showed anti-microbial potential but they showed different results against different organisms. Butanol extracted compounds showed anti-microbial potential against all MDR strains including Gram +ve and -ve strains. The results of synergism effect of all parts evaluate that, In the used concentration of *C.colocynthis* all the factors individually produced less anti- oxidant potential as compared to the synergism effect that not only shown the maximum inhibition but give maximum result in minimum concentration of 93µg/mL.

The maximum DPPH inhibitory concentration of *Citrullus Colocynthis* methanolic root extract is 23.63% alone (Ahmned, et al., 2019) while in the synergism effect with other parts including peel, seed and pulp it gave 93.75% DPPH scavenging activity with the same methanolic extract. The maximum inhibitory effect of fruit was 88.8% according to (Benariba, et al., 2013), while with synergism with its own root can produce 92.76% inhibition with the concentration of 93µg/ml indicating the synergism effect to be more effective even in the less concentration.

The Chloroform extract of *Citrullus colocynthis* fruit cannot produce antimicrobial activity according to (Priyavardhini, S., Vasantha, K., Umeadevi & M., 2009) while the study reveal all the tested strains of various pathogenic species showed sensitivity towards the same chloroform extract of the *Citrullus colocynthis* fruit, seed, peel and root and some resistant pathogens produce significant results.

The minimum inhibitory concentration of *Citrullus colocynthis* fruit chloroform extract can produce antimicrobial activity against ATCC strains of *Staphylococcus aureus* (MRSA) by MIC of 23.375µg/ml and MBC of 40.376 µg/ml and against *Pseudomonas aurignosa* with MIC of 25.375 µg/ml and MBC of 50.750 µg/ml (Belsem, et al., 2012) while the multi Drug resistant strain of *Staphylococcus aureus* (MRSA) can be inhibited by using MIC with IC50 of 150µg/ml and for *Pseudomonas aurignosa* was 300 µg/ml.

Conclusion

Resistance towards multi drugs is considered as one of the major risk for Health Care System, it is necessary to device new ways in order to prevent the spread of these pathogens.. According to findings, *Citrullus colocynthis* can be used as medicinal plant as its various compounds isolated through organic solvents showed antimicrobial activity against MDR-pathogens. Maximum activity against MDR pathogens of *Pseudomonas Aeurignosa*, *Staphylococcus aureus* (MRSA), *Klebsiella pneumoniae*, *Stenotrophomonas maltophilia*, *Salmonella typhi*, *Salmonella paratyphi*, *Acinetobacter baumannii* and *Enterococcus faecalis* was shown by Butanol extract followed by Chloroform, Acetone, Ethanol, Hexane and DMSO extract. The plant also showed 92% anti-oxidant potential in the least concentration of 93µg/mL. Suggestions

- Evaluation of antimicrobial activity of *C.colocynthis* against XDR can be include.
- Evaluation of other organic Solvents Extracts antimicrobial activity can perform.
- Evaluation of the responsible compounds and their classification as broad spectrum and narrow spectrum antibiotics.

Reference

1. Palombo, E. A., & Semple, S. J. (2001). Antibacterial activity of traditional Australian medicinal plants. *Journal of ethnopharmacology*, 77(2-3), 151-157.
2. Abd El-Ghani, M. M. (2016). Traditional medicinal plants of Nigeria: an overview. *Agriculture and Biology Journal of North America*, 7(5), 220-247.
3. MALEK, T., Chaudhari, K., & Maitrey, B. (2022). A REVIEW ON REPORTED ETHANOMEDICINAL PLANTS OF ARAVALLI DISTRICT, GUJARAT, INDIA. *VIDYA-A JOURNAL OF GUJARAT UNIVERSITY*, 1(2), 76-79.
4. Hao, D. C., & Xiao, P. G. (2015). Genomics and evolution in traditional medicinal plants: road to a healthier life. *Evolutionary Bioinformatics*, 11, EBO-S31326.
5. Alqahtani, A. S., Ullah, R., & Shahat, A. A. (2022). Bioactive constituents and toxicological evaluation of selected antidiabetic medicinal plants of Saudi Arabia. *Evidence-Based Complementary and Alternative Medicine*, 2022.
6. Anup, K., Mohan, K., Suraj, S., Sandip, F., Bhushan, F., & Prashant, W. (2010). Antimicrobial activity of some important medicinal plants of India against some plant and human pathogens. *Research Journal of Pharmacy and Technology*, 3(3), 924-926.
7. Shakya, A. K. (2016). Medicinal plants: Future source of new drugs. *International journal of herbal medicine*, 4(4), 59-64.
8. Dar, R. A., Shahnawaz, M., & Qazi, P. H. (2017). General overview of medicinal plants: A review. *The journal of phytopharmacology*, 6(6), 349-351.
9. Dar, R. A., Shahnawaz, M., & Qazi, P. H. (2017). General overview of medicinal plants: A review. *The journal of phytopharmacology*, 6(6), 349-351.
10. Mustapa, A. N., Martin, Á., Mato, R. B., & Cocero, M. J. (2015). Extraction of phytochemicals from the medicinal plant *Clinacanthus nutans* Lindau by microwave-assisted extraction and supercritical carbon dioxide extraction. *Industrial Crops and Products*, 74, 83-94.
11. Maatooq, G. T., El-Sharkawy, S. H., Afifi, M. S., & Rosazza, J. P. (1997). Cphydroxybenzoylglycoflavones from *Citrullus colocynthis*. *Phytochemistry*, 44(1), 187-190.
12. Levi, A., Thomas, C. E., Keinath, A. P., & Wehner, T. C. (2001). Genetic diversity among watermelon (*Citrullus lanatus* and *Citrullus colocynthis*) accessions. *Genetic Resources and Crop Evolution*, 48, 559-566.

13. Kumar, S., Kumar, D., Saroha, K., Singh, N., & Vashishta, B. (2008). Antioxidant and free radical scavenging potential of *Citrullus colocynthis* (L.) Schrad. methanolic fruit extract. *Acta Pharmaceutica*, 58(2), 215-220.
14. Bhasin, A., Singh, S., & Garg, R. (2020). Nutritional and medical importance of *Citrullus colocynthis*-A review. *Plant Archives*, 20(2), 3400-3406.
15. Marzouk, B., Marzouk, Z., Décor, R., Edziri, H., Haloui, E., Fenina, N., & Aouni, M. (2009). Antibacterial and anticandidal screening of Tunisian *Citrullus colocynthis* Schrad. from Medenine. *Journal of ethnopharmacology*, 125(2), 344-349.
16. Ventola, C. L. (2015). The antibiotic resistance crisis: part 1: causes and threats. *Pharmacy and therapeutics*, 40(4), 277.
17. Barghamdi, B., Ghorat, F., Asadollahi, K., Sayehmiri, K., Peyghambari, R., & Abangah, G. (2016). Therapeutic effects of *Citrullus colocynthis* fruit in patients with type II diabetes: A clinical trial study. *Journal of pharmacy & bioallied sciences*, 8(2), 130.
18. Rabizadeh, F., Mirian, M. S., Doosti, R., Kiani-Anbouhi, R., & Eftekhari, E. (2022). Phytochemical Classification of Medicinal Plants Used in the Treatment of Kidney Disease Based on Traditional Persian Medicine. *Evidence-Based Complementary and Alternative Medicine*, 2022.
19. Achilonu, M., Shale, K., Arthur, G., Naidoo, K., & Mbatha, M. (2018). Phytochemical benefits of agroresidues as alternative nutritive dietary resource for pig and poultry farming. *Journal of Chemistry*, 2018, 1-15.
20. Mariod, A. A., & Jarret, R. L. (2022). Antioxidant, antimicrobial, and antidiabetic activities of *Citrullus colocynthis* seed oil. In *Multiple Biological Activities of Unconventional Seed Oils* (pp. 139-146). Academic Press.
21. Eidi, S., Azadi, H. G., Rahbar, N., & Mehmannaavaz, H. R. (2015). Evaluation of antifungal activity of hydroalcoholic extracts of *Citrullus colocynthis* fruit. *Journal of herbal medicine*, 5(1), 36-40.
22. da Silva, J. A. T., & Hussain, A. I. (2017). *Citrullus colocynthis* (L.) Schrad.(colocynth): Biotechnological perspectives. *Emirates Journal of Food and Agriculture*, 83-90.
23. Dabe, N. E., & Kefale, A. T. (2017). Antidiabetic effects of *Artemisia* species: a systematic review. *Ancient science of life*, 36(4), 175.
24. Finch, C. E. (2005). Developmental origins of aging in brain and blood vessels: an overview. *Neurobiology of aging*, 26(3), 281-291.
25. Duke, J. A. (2008). *Duke's handbook of medicinal plants of Latin America*. CRC press.
26. Alhawiti, N. M. (2018). Antiplatelets and profibrinolytic activity of *Citrullus colocynthis* in control and high-fat diet-induced obese rats: mechanisms of action. *Archives of physiology and biochemistry*, 124(2), 156-166.
27. Ballotin, V. R., Bigarella, L. G., de Mello Brandão, A. B., Balbinot, R. A., Balbinot, S. S., & Soldera, J. (2021). Herb-induced liver injury: Systematic review and meta-analysis. *World Journal of Clinical Cases*, 9(20), 5490.
28. Sanadgol, N., Najafi, S., Ghasemi, L. V., Motalleb, G., & Estakhr, J. (2011). A study of the inhibitory effects of *Citrullus colocynthis* (CCT) using hydro-alcoholic extract on the expression of cytokines: TNF- α and IL-6 in high fat diet-fed mice towards a cure for diabetes mellitus. *Journal of pharmacognosy and phytotherapy*, 3(6), 81-88.
29. Pashmforosh, M., Rajabi Vardanjani, H., Rajabi Vardanjani, H., Pashmforosh, M., & Khodayar, M. J. (2018). Topical anti-inflammatory and analgesic activities of *Citrullus colocynthis* extract cream in rats. *Medicina*, 54(4), 51.
30. Tannin-Spitz, T., Grossman, S., Dovrat, S., Gottlieb, H. E., & Bergman, M. (2007). Growth inhibitory activity of cucurbitacin glucosides isolated from *Citrullus colocynthis* on human breast cancer cells. *Biochemical pharmacology*, 73(1), 56-67.

Hafiza Bisma Nadeem: Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains

31. Rizvi, T. S., Khan, A. L., Ali, L., Al-Mawali, N., Mabood, F., Hussain, J., ... & Al-Harrasi, A. (2018). In vitro oxidative stress regulatory potential of *Citrullus colocynthis* and *Tephrosia apollinea*. *Acta Pharmaceutica*, 68(2), 235-242.
32. Gupta, S. C., Tripathi, T., Paswan, S. K., Agarwal, A. G., Rao, C. V., & Sidhu, O. P. (2018). Phytochemical investigation, antioxidant and wound healing activities of *Citrullus colocynthis* (bitter apple). *Asian Pacific Journal of Tropical Biomedicine*, 8(8), 418.
33. Mariod, A. A., & Jarret, R. L. (2022). Antioxidant, antimicrobial, and antidiabetic activities of *Citrullus colocynthis* seed oil. In *Multiple Biological Activities of Unconventional Seed Oils* (pp. 139-146). Academic Press.
34. Afzal, M., Khan, A. S., Zeshan, B., Riaz, M., Ejaz, U., Saleem, A., ... & Ahmed, N. (2023). Characterization of bioactive compounds and novel proteins derived from promising source *Citrullus colocynthis* along with in-vitro and in-vivo activities. *Molecules*, 28(4), 1743.
35. Javadzadeh, H. R., Davoudi, A., Davoudi, F., Valizadegan, G., Goodarzi, H., Mahmoodi, S., ... & Faraji, M. (2013). *Citrullus colocynthis* as the Cause of Acute Rectorrhagia. *Case reports in emergency medicine*, 2013.
36. Al-Nablsi, S., El-Keblawy, A., Ali, M. A., Mosa, K. A., Hamoda, A. M., Shanableh, A., ... & Soliman, S. S. (2022). Phenolic contents and antioxidant activity of *Citrullus Colocynthis* fruits, growing in the hot arid desert of the UAE, influenced by the fruit parts, accessions, and seasons of fruit collection. *Antioxidants*, 11(4), 656.
37. Rizvi, T. S., Mabood, F., Ali, L., Al-Broumi, M., Al Rabani, H. K., Hussain, J., ... & Al-Harrasi, A. (2018). Application of NIR spectroscopy coupled with PLS regression for quantification of total polyphenol contents from the fruit and aerial parts of *Citrullus colocynthis*. *Phytochemical Analysis*, 29(1), 16-22.
38. Lemos, M. F., Lemos, M. F., Pacheco, H. P., Guimarães, A. C., Fronza, M., Endringer, D. C., & Scherer, R. (2017). Seasonal variation affects the composition and antibacterial and antioxidant activities of *Thymus vulgaris*. *Industrial Crops and Products*, 95, 543-548.
39. Birgani, G. A., Ahangarpour, A., Khorsandi, L., & Moghaddam, H. F. (2018). Anti-diabetic effect of betulinic acid on streptozotocin-nicotinamide induced diabetic male mouse model. *Brazilian Journal of Pharmaceutical Sciences*, 54.
40. Patel, D. K., Prasad, S. K., Kumar, R., & Hemalatha, S. (2012). An overview on antidiabetic medicinal plants having insulin mimetic property. *Asian Pacific journal of tropical biomedicine*, 2(4), 320-330.
41. Chauhan, A., Semwal, D. K., Mishra, S. P., & Semwal, R. B. (2015). Ayurvedic research and methodology: Present status and future strategies. *Ayu*, 36(4), 364.
42. Shaikh, S. I., Nagarekha, D., Hegade, G., & Marutheesh, M. (2016). Postoperative nausea and vomiting: A simple yet complex problem. *Anesthesia, essays and researches*, 10(3), 388.
43. Koche, D., Shirsat, R., & Kawale, M. A. H. E. S. H. (2016). An overerview of major classes of phytochemicals: their types and role in disease prevention. *Hislopia Journal*, 9(1/2), 0976-2124.
44. Mohiuddin, A. K. (2019). A brief review of traditional plants as sources of pharmacological interests. *Open Journal of Plant Science*, 4(1), 1-8.
45. Kafshgari, H. S., Yazdanian, M., Ranjbar, R., Tahmasebi, E., Mirsaeed, S. R. G., Tebyanian, H., ... & Goli, H. R. (2019). The effect of *Citrullus colocynthis* extracts on *Streptococcus mutans*, *Candida albicans*, normal gingival fibroblast and breast cancer cells. *Journal of Biological Research Bollettino della Società Italiana di Biologia Sperimentale*, 92(1).
46. Abdulridha, M. K., Al-Marzoqi, A. H., & Ghasemian, A. (2020). The anticancer efficiency of *Citrullus colocynthis* toward the colorectal cancer therapy. *Journal of Gastrointestinal Cancer*, 51, 439-444
47. Lahfa, F. B., Azzi, R., Mezouar, D., & Djaziri, R. (2017). Hypoglycemic effect of *Citrullus colocynthis* extracts. *Phytothérapie*, 15(2), 50-56.

48. Al-Hwaiti, M. S., Alsbou, E. M., Abu Sheikha, G., Bakchiche, B., Pham, T. H., Thomas, R. H., & Bardaweel, S. K. (2021). Evaluation of the anticancer activity and fatty acids composition of “Handal”(Citrullus colocynthis L.) seed oil, a desert plant from south Jordan. *Food Science & Nutrition*, 9(1), 282-289.
49. Rajizadeh, M. A., Aminizadeh, A. H., Esmailpour, K., Bejeshk, M. A., Sadeghi, A., & Salimi, F. (2023). Investigating the effects of Citrullus colocynthis on cognitive performance and anxiety-like behaviors in STZ-induced diabetic rats. *International Journal of Neuroscience*, 133(4), 343-355
50. Rahbar, A. R., & Nabipour, I. (2010). The hypolipidemic effect of Citrullus colocynthis on patients with hyperlipidemia. *Pakistan journal of biological sciences: PJBS*, 13(24), 1202-1207.
51. Daradka, H., Almasad, M. M., WSh, Q., El-Banna, N. M., & Samara, O. H. (2007). Hypolipidaemic effects of Citrullus colocynthis L. in rabbits. *Pakistan journal of biological sciences: PJBS*, 10(16), 2768-2771.
52. Jeon, J. H., & Lee, H. S. (2014). Biofunctional constituent isolated from Citrullus colocynthis fruits and structure–activity relationships of its analogues show acaricidal and insecticidal efficacy. *Journal of agricultural and food chemistry*, 62(34), 8663-8667.
53. Rahuman, A. A., Venkatesan, P., & Gopalakrishnan, G. (2008). Mosquito larvicidal activity of oleic and linoleic acids isolated from Citrullus colocynthis (Linn.) Schrad. *Parasitology research*, 103, 1383-1390.
54. Chaweche, R., Njeh, F., Hamed, N., Damak, M., Ayadi, A., Hammami, H., & Mezghani-Jarraya, R. (2017). A study of the molluscicidal and larvicidal activities of Citrullus colocynthis (L.) leaf extract and its main cucurbitacins against the mollusc Galba truncatula, intermediate host of Fasciola hepatica. *Pest management science*, 73(7), 1473-1477.
55. Meybodi, M. S. K. (2020). A review on pharmacological activities of Citrullus colocynthis (L.) Schrad. *Asian J. Res. Rep. Endocrinol*, 25, 25-34.
56. da Silva, J. A. T., & Hussain, A. I. (2017). Citrullus colocynthis (L.) Schrad.(colocynth): Biotechnological perspectives. *Emirates Journal of Food and Agriculture*, 83-90.
57. Amamou, F., Bouafia, M., Chabane-Sari, D., Meziane, R. K., & Nani, A. (2011). Citrullus colocynthis: a desert plant native in Algeria, effects of fixed oil on blood homeostasis in Wistar rat. *Journal of Natural Product and Plant Resources*, 1, 1-7.
58. Savithramma, N., Sulochana, C., & Rao, K. N. (2007). Ethnobotanical survey of plants used to treat asthma in Andhra Pradesh, India. *Journal of Ethnopharmacology*, 113(1), 54-61.
59. Bouldin, A. S., Smith, M. C., Garner, D. D., Szeinbach, S. L., Frate, D. A., & Croom, E. M. (1999). Pharmacy and herbal medicine in the US. *Social science & medicine*, 49(2), 279-289.
60. Lavie, D., Willner, D., & Merenlender, Z. (1964). Constituents of Citrullus colocynthis (L.) Schrad. *Phytochemistry*, 3(1), 51-56.
61. Meybodi, M. S. K. (2020). A review on pharmacological activities of Citrullus colocynthis (L.) Schrad. *Asian J. Res. Rep. Endocrinol*, 25, 25-34.
62. Rajizadeh, M. A., Aminizadeh, A. H., Esmailpour, K., Bejeshk, M. A., Sadeghi, A., & Salimi, F. (2023). Investigating the effects of Citrullus colocynthis on cognitive performance and anxiety-like behaviors in STZ-induced diabetic rats. *International Journal of Neuroscience*, 133(4), 343-355.
63. Mohiuddin, A. K. (2019). A brief review of traditional plants as sources of pharmacological interests. *Open Journal of Plant Science*, 4(1), 1-8.
64. Roy, R. K., Thakur, M., & Dixit, V. K. (2007). Effect of citrullus colocynthis. On hair growth in albino rats. *Pharmaceutical biology*, 45(10), 739-744.
65. Chaturvedi, M., Mali, P. C., & Ansari, A. S. (2003). Induction of reversible antifertility with a crude ethanol extract of Citrullus colocynthis Schrad fruit in male rats. *Pharmacology*, 68(1), 38-48.
66. Kumar, D., Kumar, A., & Prakash, O. (2012). Potential antifertility agents from plants: A comprehensive review. *Journal of Ethnopharmacology*, 140(1), 1-32.

Hafiza Bisma Nadeem: Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains

67. Sharma, A., Sharma, P., Chaturvedi, M., & Joshi, S. C. (2014). Effect of *Citrullus colocynthis* on function of cauda epididymis and accessory reproductive organs of male rats. *World Journal of Pharmaceutical Research*, 3(2), 2406-2419.
68. Vivas, R., Barbosa, A. A. T., Dolabela, S. S., & Jain, S. (2019). Multidrug-resistant bacteria and alternative methods to control them: an overview. *Microbial Drug Resistance*, 25(6), 890-908.
69. Medina, E., & Pieper, D. H. (2016). Tackling threats and future problems of multidrug-resistant bacteria. How to overcome the antibiotic crisis: facts, challenges, technologies and future perspectives, 3-33.
70. Basak, S., Singh, P., & Rajurkar, M. (2016). Multidrug resistant and extensively drug resistant bacteria: a study. *Journal of pathogens*, 2016.
71. Sharma, A. (2011). Antimicrobial resistance: no action today, no cure tomorrow. *Indian Journal of Medical Microbiology*, 29(2), 91.
72. Cohen, M. L. (2000). Changing patterns of infectious disease. *Nature*, 406(6797), 762-767.
73. Oliver, A., Mulet, X., López-Causapé, C., & Juan, C. (2015). The increasing threat of *Pseudomonas aeruginosa* high-risk clones. *Drug Resistance Updates*, 21, 41-59.
74. Reynolds, D., & Kollef, M. (2021). The epidemiology and pathogenesis and treatment of *Pseudomonas aeruginosa* infections: an update. *Drugs*, 81(18), 2117-2131.
75. Poole, K. (2011). *Pseudomonas aeruginosa*: resistance to the max. *Frontiers in microbiology*, 2, 65.
76. Breidenstein, E. B., de la Fuente-Núñez, C., & Hancock, R. E. (2011). *Pseudomonas aeruginosa*: all roads lead to resistance. *Trends in microbiology*, 19(8), 419-426.
77. Oliver, A., Mulet, X., López-Causapé, C., & Juan, C. (2015). The increasing threat of *Pseudomonas aeruginosa* high-risk clones. *Drug Resistance Updates*, 21, 41-59.
78. Horcajada, J. P., Montero, M., Oliver, A., Sorlí, L., Luque, S., Gómez-Zorrilla, S., ... & Grau, S. (2019). Epidemiology and treatment of multidrug-resistant and extensively drug-resistant *Pseudomonas aeruginosa* infections. *Clinical microbiology reviews*, 32(4), e00031-19.
79. Hua, X., Liu, L., Fang, Y., Shi, Q., Li, X., Chen, Q., ... & Yu, Y. (2017). Colistin resistance in *Acinetobacter baumannii* MDR-ZJ06 revealed by a multiomics approach. *Frontiers in cellular and infection microbiology*, 7, 45.
80. Ni, W., Han, Y., Zhao, J., Wei, C., Cui, J., Wang, R., & Liu, Y. (2016). Tigecycline treatment experience against multidrug-resistant *Acinetobacter baumannii* infections: a systematic review and meta-analysis. *International journal of antimicrobial agents*, 47(2), 107-116.
81. Adegoke, A. A., Stenström, T. A., & Okoh, A. I. (2017). *Stenotrophomonas maltophilia* as an emerging ubiquitous pathogen: looking beyond contemporary antibiotic therapy. *Frontiers in microbiology*, 8, 2276.
82. Chang, Y. T., Lin, C. Y., Chen, Y. H., & Hsueh, P. R. (2015). Update on infections caused by *Stenotrophomonas maltophilia* with particular attention to resistance mechanisms and therapeutic options. *Frontiers in microbiology*, 6, 893.

83. Nguyen, T. H. T., Nguyen, N. A. T., Nguyen, H. D., Nguyen, T. T. H., Le, M. H., Pham, M. Q., ... & Pham, H. N. (2023). Plant Secondary Metabolites on Efflux-Mediated Antibiotic Resistant *Stenotrophomonas Maltophilia*: Potential of Herbal-Derived Efflux Pump Inhibitors. *Antibiotics*, 12(2), 421.
84. Dadashi, M., Hajikhani, B., Nazarinejad, N., Nourisepehr, N., Yazdani, S., Hashemi, A., ... & Sameni, F. (2023). Global prevalence and distribution of antibiotic resistance among clinical isolates of *Stenotrophomonas maltophilia*: a systematic review and meta-analysis. *Journal of Global Antimicrobial Resistance*.
85. Kumar, G., & Tudu, A. K. (2023). Tackling multidrug-resistant *Staphylococcus aureus* by natural products and their analogues acting as NorA efflux pump inhibitors. *Bioorganic & Medicinal Chemistry*, 117187.
86. Kime, L., Waring, T., Mohamad, M., Mann, B. F., & O'Neill, A. J. (2023). Resistance to antibacterial antifolates in multidrug-resistant *Staphylococcus aureus*: prevalence estimates and genetic basis. *Journal of Antimicrobial Chemotherapy*, 78(5), 1201-1210.
87. Chand, U., Priyambada, P., & Kushawaha, P. K. (2023). *Staphylococcus aureus* vaccine strategy: promise and challenges. *Microbiological Research*, 127362.
88. Al-Trad, E. A. I., Che Hamzah, A. M., Puah, S. M., Chua, K. H., Hanifah, M. Z., Ayub, Q., ... & Yeo, C. C. (2023). Complete Genome Sequence and Analysis of a ST573 Multidrug-Resistant Methicillin-Resistant *Staphylococcus aureus* SauR3 Clinical Isolate from Terengganu, Malaysia. *Pathogens*, 12(3), 502.
89. Hussain, T., Shami, A., Rafiq, N., Khan, S., Kabir, M., Khan, N. U., ... & Usman, T. (2023). Antimicrobial Usage and Detection of Multidrug-Resistant *Staphylococcus aureus*: Methicillin-and Tetracycline-Resistant Strains in Raw Milk of Lactating Dairy Cattle. *Antibiotics*, 12(4), 673.
90. Aljasir, S. F., & D'Amico, D. J. (2023). Anti-infective properties of the protective culture *Hafnia alvei* B16 in food and intestinal models against multi-drug resistant *Salmonella*. *Food Microbiology*, 110, 104159.
91. Mina, S. A., Hasan, M. Z., Hossain, A. Z., Barua, A., Mirjada, M. R., & Chowdhury, A. M. A. (2023). The Prevalence of Multi-Drug Resistant *Salmonella typhi* Isolated From Blood Sample. *Microbiology Insights*, 16, 11786361221150760.
92. Chou, S. H., Wan, T. W., Shiau, C. W., Chen, L. H., Lin, H. C., & Chiu, H. C. (2023). Repurposing the Tyrosine Kinase Inhibitor Nilotinib for Use Against Intracellular Multidrug-Resistant *Salmonella Typhimurium*. *Journal of Microbiology, Immunology and Infection*.
93. Igbinosa, I. H., Amolo, C. N., Beshiru, A., Akinnibosun, O., Ogofure, A. G., El-Ashker, M., ... & Igbinosa, E. O. (2023). Identification and characterization of MDR virulent *Salmonella* spp isolated from smallholder poultry production environment in Edo and Delta States, Nigeria. *Plos one*, 18(2), e0281329.

Hafiza Bisma Nadeem: Evaluation of Anti-Oxidant Potential and Novel Synergism Effect of *Citrullus colocynthis* against MDR pathogenic Strains

94. García, P., Moscoso, M., Fuentes-Valverde, V., Rodicio, M. R., Herrera-León, S., & Bou, G. (2023). A highly-safe live auxotrophic vaccine protecting against disease caused by non-typhoidal *Salmonella* Typhimurium in mice. *Journal of Microbiology, Immunology and Infection*, 56(2), 324336.
95. Pereira-Dias, J., Taneja, N., Mahindroo, J., Maheshwari, G., Patel, P. J., Thu, T. N. H., ... & Mylona, E. (2023). The genomic characterization of *Salmonella* Paratyphi A from an outbreak of enteric fever in Vadodara, India. *Microbial Genomics*, 9(1), 000914.
96. Sajib, M. S., Tanmoy, A. M., Hooda, Y., Rahman, H., Munira, S. J., Sarkar, A., ... & Saha, S. (2023). 23-year trends indicate low rates antimicrobial resistance in *Salmonella* Paratyphi A. medRxiv, 202302.
97. Mahapatra, S. R., Dey, J., Kushwaha, G. S., Puhan, P., Mohakud, N. K., Panda, S. K., ... & Suar, M. (2022). Immunoinformatic approach employing modeling and simulation to design a novel vaccine construct targeting MDR efflux pumps to confer wide protection against typhoidal *Salmonella* serovars. *Journal of Biomolecular Structure and Dynamics*, 40(22), 11809-11821.
98. Umair, M., & Siddiqui, S. A. (2020). Antibiotic susceptibility patterns of *Salmonella typhi* and *Salmonella paratyphi* in a tertiary care hospital in Islamabad. *Cureus*, 12(9).
99. Bassetti, M., Righi, E., Carnelutti, A., Graziano, E., & Russo, A. (2018). Multidrug-resistant *Klebsiella pneumoniae*: challenges for treatment, prevention and infection control. *Expert review of anti-infective therapy*, 16(10), 749-761.
100. Tang, M., Kong, X., Hao, J., & Liu, J. (2020). Epidemiological characteristics and formation mechanisms of multidrug-resistant hypervirulent *Klebsiella pneumoniae*. *Frontiers in microbiology*, 11, 581543.
101. Lin, Z. W., Zheng, J. X., Bai, B., Xu, G. J., Lin, F. J., Chen, Z., ... & Deng, Q. W. (2020). Characteristics of hypervirulent *Klebsiella pneumoniae*: does low expression of *rmpA* contribute to the absence of hypervirulence?. *Frontiers in microbiology*, 11, 436.
102. van Harten, R. M., Willems, R. J., Martin, N. I., & Hendrickx, A. P. (2017). Multidrug-resistant enterococcal infections: new compounds, novel antimicrobial therapies?. *Trends in microbiology*, 25(6), 467-479.
103. Adesida, S. A., Ezenta, C. C., Adagbada, A. O., Aladesokan, A. A., & Coker, A. O. (2017). Carriage of multidrug resistant *Enterococcus faecium* and *Enterococcus faecalis* among apparently healthy humans. *African journal of infectious diseases*, 11(2), 83-89.
104. Bhatt, P., Patel, A., Sahni, A. K., Praharaj, A. K., Grover, N., Chaudhari, C. N., ... & Kulkarni, M. (2015). Emergence of multidrug resistant enterococci at a tertiary care centre. *medical journal armed forces india*, 71(2), 139-144.

105. Freitas, A. R., Pereira, A. P., Novais, C., & Peixe, L. (2021). Multidrug-resistant high-risk *Enterococcus faecium* clones: can we really define them?. *International journal of antimicrobial agents*, 57(1), 106227.