Molecular Detection for Some Virulence Factors of Salmonella typhi from Iraqi Patients

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Received: 11 May 2023	Accepted: 10 June 2023
Citation: Abdul-Khadum	n MAR, Saleem AJ, Karim AZ (2023) Molecular Detection for Some Virulence Factors of
Salmonella typhi from Iraq	i Patients. History of Medicine 9(2): 647–653. https://doi.org/10.17720/2409-5834.v9.2.2023.081

Abstract

In current study, 29 clinical isolates Salmonella typhi were collected from different Private laboratories in Diyala. In addition of phenotypic method, all isolates diagnosed as S. typhi by genotypic method was done by Polymerase Chain Reaction (PCR) technique to detection 16SrRNA gene. And detection of adhesion factors associated genes that encoding for cdtB, tviA, fimA and orfL by using specific primers which showed the prevalence in percentage 100% for all genes.

Keywords

Salmoella typhi ,16SrRNA, cdtB, tviA, fimA and orfL genes, Virulence Factors genes.

The genus salmonella is one of the genera of the intestinal bacterial family and includes about 2600 serotypes, salmonella is a gram stain-negative bacterium, classified into two types S. enterica and S. bongori (ALJobouri, 2019). They move by peritrichous flagella, are non-spore-forming , contain pilli (Grimont et al., 2000). Salmonella grows at optimum pH between 6,5-7.5 and temperature is 37C, also it can grow at pH between 3-9.5 and temperature between 3-52C(ALJobouri, 2019).

Possessing special Salmonella invasion proteins such as SipB and SipA these proteins cause death of phagocytic cells . Many genes of virulence factors for salmonella are clustered in certain chromosomal regions known as Salmonella pathogenicity islands (SPIs) (Santos et al.,2003).SPIs are chromosomal site containing virulence genes ,it's found only pathogenic bacteria and it's located either on chromosomes or plasmid surrounded by repeated sequences and differs in contain of G/C content in relation to the surrounding area (AL-Muhannak ,2020).

Recent international statistics indicate that typhoid fever contributes to 21.6-26.9 million cases of illness and 21,600 deaths annually (WHO, 2018; Wong et al., 2019).

S.typhi has combination of properties that make it an effective pathogen, as it has many Virulence factors ,that play an important role at different stages of infection ,some of which are necessary for entry and survival inside and outside cells, including H and Vi antigens ,Lipopolysaccharide ,endotoxin and siderophores (Abbas,2012).

tviA gene encodes a regulatory protein that plays an important role in regulating the expression of antigen Vi ,flagella, and a number of genes necessary for host invasion (Santander et al., 2008). This gene located on pathogenicity island of salmonella SPI-7, which has a capacity of 134Mb. The synthesis and transport proteins of the Vi capsular polysaccharide of S.typhi are encoded by viaB Operon (Zhang et al.,2018).

cdtB gene is one of the most important newly known invasion factors for S.typhi, which represents a completely wide range of other intestinal pathogens , cytotoxicity entails the so-called DNA Damage response (DDR), which is characterized by irreparable cell damage and, consequently, cell death by apoptosis (AL-Oqaili, 2019).Typhoid toxin (A2B5) contains two active subunits connected by a disulfide bridge (cdtB and pltA) connected with five subunits (pentameric pltB) of auxiliary memory aid - memory, secreted from S.typhi only an intracellular and intracellular vacuole containing salmonella (SCV) (Song et al., 2013).

fimA gene encodes the main unit of the first type of helminth , there is a significant similarity in the nucleotide sequence of the fimA gene of E.coli and Klebsiella pneumonia but they are completely heterogeneous(COHEN,1996). The genes fimA, fimI, fimC, fimD, fimH, fimF are assembled in the form of a single operon located under the control of the catalyst fimA encodes for a protein involved in biosynthesis and the synthesis of aliphates (Kolenda, 2019). orfL gene was diagnosed in Salmonella Pathogenicity Island 4 (SPI-4) and is involved in adhesion, spontaneous transport and colonization. orfL gene is essential for survival inside macrophages and possibly carries a system involved in the excretion of toxins (LEGBA et al., 2017).

Methods And Materials

Sample collection and identification

During the period from September 2021 to December 2021, 75 blood specimens were collected at suspected outpatient's patients for typhoid fever in Diyala .The specimens were transplanted on MacConkey ,Blood agars also on XLD and S-S agars ,that incubated at temperature of 37c for 24-48 hours, diagnosed on phenotypic, cultural characteristics ,microscopy with gram stain and biochemical tests: catalase and imvic (Macfaddin ,2000), as well as Molecular detection by 16SrRNA gene.

Molecular Identification

DNA extraction from *S.typhi* isolates by Genomic DNA extraction kit (ABIO, USA). DNA preparation was then analyzed via electrophoresis thereof 1.5% agarose gel. PCR be used to amplify the entire sequences of the genes studied in this research. The specific primers (Macrogen, Korea) utilized for the expansion to these genes were shown in (table1).

PCR mixture was prepared by adding 12.5 μ l of GoTaq®Green master Mix (2X) promega,2 μ l template DNA, 1 μ l from each forward and reverse primers with final concentration 1 poml/ μ l, finally volume was completed to 25 μ l by adding nuclease free water. PCR condition illustrated in Table (2), stained with ethidium bromide and visualized by transilluminator.

Gene	Primer sequences (5'- 3')	size(bp)	Reference	
16SrRNA	F:CCTGGACAAAGACTGACGCT	523	Al- Oqaili,2019	
	R: CGCTTCTCTTTGTATGCGCC			
fimA	F: GTGAGCGGCGGTACTATTC	451		
	R: TAAAGGTGGCGTCGGCATT			
cdtB	F:TAAGTGGTACTGCCGGTGTG	508		
	R:GTAGGTGCGAGTACGGCTAC			
tviA	F: GTTATTTCAGCATAAGGAG	599	Liaquat et al. 2018)(
	R: ACTTGTCCGTGTTTTACTC			
orfL	F: GGAGTATCGATAAAGATGTT	323	(Albanwawy and Abdul-Lateef,2021)	
	R: GCGCGTAACGTCAGAATCA			

Table (1): Primers and amplified PCR products used in study

Amplified gene	Initial denaturation	No.of cycles	Denaturation	Annealing	Elongation	Final extension
16SrRNA	5min. at 95C	30	30sec.at95C°	30sec.at58C°	$30 \text{sec.} 72 \text{C}^{\circ}$	7min.at72C
fimA	5min.at95C°	30	30sec.at95C°	30sec.at57C°	30sec.72C°	7min.at72C
cdtB	5min.at 95C°	30	30sec.at95C°	30sec.at58C°	30sec.72C°	7min.at72C
TviA	5min.at 95C°	30	30sec.at95C°	30sec.at 55C	30sec.72C°	7min.at72C
orfL	5min.at 95C°	30	30sec.at95C°	30sec.at58C°	30sec.72C°	7min.at72C

Table (2): PCR condition to genes used in study

Results and Discussions

Identification of S. typhi

Bacteria isolates have been diagnosed in two ways: the first is phenotypic by microscopic ,it's gram negative and cultural characteristics (on XLD agar showed red colorwith or without black center for H_2S production ,and on S-S agar showed pale yellow for not fermentation lactose with a black center for producing H_2S), in addition of biochemical tests, the second is genotypic by polymerase chain reaction (PCR).

The results of the current study showed 29 (38%) isolation of *S.typhi* depending on phenotypic and culture characteristics. Also genetically diagnosed by *16SrRNA* gene and the isolates of *S.typhi* possessed the *16S rRNA* gene 100%. (Fig.1).



Fig.1: Agaros gel electrophoresis (1.5 agaros,100 V for 1h,for 16SrRNA gene, DNA ladder (100-1500 pb).

This gene contains conserved regions that overlap with variable regions , there are 9 variable regions, that are useful in determining genus and orders bacteria because 16SrRNA is present in all bacterial species and also has little variable(Suardana,2014;Srinivasan et al ,2015).

The results of our study some virulence factors *orfL*, *cdtB*, *tviA* and *fimA genes*, it was found that isolates carried all genes rate 100%.

orfL gene is encoded into the adhesion protein , which helps in the adhesion and survival of bacteria inside phagocytic cells, and the bacteria also have a secretion mechanism that regulates the secretion of toxins by stimulating the programmed death of host cells and thus promotes the pathogenicity of *S.typhi* (Gassama-sow et al.,2006; Ramatle et al.,2022).Fig.2



Fig.2: Agaros gel electrophoresis (1.5 agaros, 100 V for 1h, for orfL gene, DNA ladder (100-1500 pb).

The results of *cdtB* gene were completely identical to the results of the study of researcher AL-Oqaili (2019) in Babylon and researcher Liaquat et al. (2018) in Pakistan. While the results of the study of researcher Al-Khafaji (2022) in AL- Najaf were not consistent with the results of the current study, where the presence of the gene was 57%. The *cdtB* gene encodes the cytolytic distending toxin protein subunit B, which contains 269 amino acids (NCBI-a). It is believed that typhoid poison contributes to the development of chronic typhoid fever and causes the appearance of symptoms (Al-Khafaji , 2022). Studies have indicated that typhoid toxin is responsible for the onset of symptoms and the transformation of the infection from acute to chronic (Johnson et al., 2018). Host cell death is due to the presence of typhoid toxin , which can be encoded by the *cdtB* gene (Al-Khafaji ,2022).Fig.3



Fig.3: Agaros gel electrophoresis (1.5 agaros, 100 V for 1h, for *cdtB* gene, DNA ladder (100-1500 pb).

Compared to the results of Al-Oqaili's (2019) study in Babylon, she showed that the presence of the *tviA* gene in *S. typhi* was 100%, so it matches the results of the current study, while the results of the study of researcher Al-Khafaji (2022) in AL- Najaf, where the percentage of the presence of the gene was 42.7%, were not consistent with the results of the study. In another study by Baker et al. (2005) and Liaquat et al. (2018) in Pakistan, where the presence of this gene was 15% and 89%, respectively, where the results were not consistent with the results of the current study..Fig.4



Fig.4: Agaros gel electrophoresis (1.5 agaros, 100 V for 1h, for tviA gene, DNA ladder (100-1500 pb)

The Vi Capsular polysaccharide of salmonella bacteria, which is the cause of typhoid infection, is important in the occurrence of infection and virulence, its biosynthesis mechanism is encoded within the ViaB site, as it consists of 10 genes involved in the expression of the *tviA* gene, polymer construction (tviB-tviE) and localization of the cell surface of the VexA-VexE capsule (Albanwawy and Abdul-Lateef,2021). The *tviA* gene encodes a capsule synthesis regulatory protein of 179 amino acids (NCBI-b).

The results of *fimA* Al-Oqaili's (2019) study in Babylon did not correspond to the results, as the presence of the gene was 84.21%. The *fimA* gene encodes for the filament type I protein type - 1 fimbrial protein which contains 184 amino acids(NCBI-c). Cilia or fimbriae filaments are located on the surface of the S-cell. typhi play an important role in the process of adhesion and invasion of host cells (Khazaal, 2019).Fig.5



Fig.5: Agaros gel electrophoresis (1.5 agaros, 100 V for 1h, for *fimA* gene, DNA ladder (100-1500 pb).

Conclusion

The local isolates in the study possess all the genes of Virulence Factors that increase the pathogenicity of bacteria ,and these genes encode adhesion proteins that are associated with the process of Biofilm formation ,as adhesion is the first stage of biofilm formation ,thus increasing the pathogenicity of bacteria through resistance to the host body's immune defenses and resistance to antibiotics.

References

- MacFaddin, J.F. (2000). Biochemical Tests for Identification of Medical Bacteria. 3rd ed. Lippincott Williams and Wilkins, USA.
- ALJobouri, Abdul Aziz T.A.F(2019).Molecular Characterization of Some Virulence Genes of Salmonella enterica in Babylon province ,Thesis M.S.C, Medicine, University of Babylon.
- Abbas, Hussein Hafidh (2012). Detection of Salmonella enterica serovar typhi in patients with acute and chronic typhoid by some bacteriological, immunological and PCR technique, Thesis M.S.C, College of Science, University of Baghdad.
- Johnson, R. (2018). Investigating the globally dominant Salmonella Typhi haplotype H58–an insight into the pathogenesis of typhoid fever.
- AL-Oqaili, NAD (2019), Detection of some virulence factors of Salmonella typhi isolated from patients' blood by PCR and phylogenetic tree, Periodicals of Engineering and Natural Sciences, 7(4) pp. 1915–1923. doi: 10.21533/pen.v7i4.941.
- LEGBA, B.,Dougnon, T. V., DEGUENON, E., HOUNMANOU, G., AGBANKPE, J., AMADOU, A., ... & DOUGNON, T. J. (2017). Pathogenicity, epidemiology and virulence factors of Salmonella species: A review. Notulae Scientia Biologicae, 9(4), 460-466.
- Cohen, H. J., Mechanda, S. M., & Lin, W. (1996). PCR amplification of the fimA gene sequence of Salmonella typhimurium, a specific method for detection of Salmonella spp. Applied and environmental microbiology, 62(12), 4303-4308.
- Khazaal, Saba Saadoon(2019). Detection Flagellin Gene of Enteric Isolates of Salmonella enterica serovar Typhi Using Conventional PCR Technique, Al-Mustansiriyah Journal of Science,30(3): ISSN: 1814-635X (print), ISSN:2521-3520 (online).
- Albanwawy, Jihad N. Abid; Abdul-Lateef, , Lamees A.(2021).Molecular Detection of Some of the Salmonella Typhi Virulence Genes Isolated in the Province of Babylon/Iraq. Annals of R.S.C.B., ISSN: 1583-6258,25(2). 675 – 685.
- Baker, S.; Sarwar, Y.; Aziz, H.; Haque, A.; Ali, A.; Dougan, G.; Wain, J. and Haque, A. (2005). Detection of Vi-negative Salmonella enterica serovar typhi in the peripheral blood of patients with typhoid fever in the Faisalabad region of Pakistan. J Clin Microbiol .43(9):4418-25. doi: 10.1128/JCM.43.9.4418-4425.2005. PMID: 16145086; PMCID: PMC1234127.

- Liaquat, S. et al. (2018) 'Virulotyping of Salmonella enterica serovar Typhi isolates from Pakistan: Absence of complete SPI-10 in Vi negative isolates', PLoS Neglected Tropical Diseases, 12(11), pp. 1–20. doi: 10.1371/journal.pntd.0006839.
- AL-Khafaji, Angham Najah (2022). Occurrences and Virulence Genes Detection of Salmonella Typhi Isolates Among Patients with Typhoid Fever. Al-Furat Al-Awsat Technical University. <u>https://orcid.org/0000-0002-4413-1905</u>.
- Gassama-Sow, A. et al. (2006) 'Characterization of virulence factors in the newly described Salmonella enterica serotype Keurmassar emerging in Senegal (sub-Saharan Africa)', Epidemiology and Infection, 134(4), pp. 741–743. doi: 10.1017/S0950268805005807.
- Ramatla,T. A.; Mphuthi, N.; Ramaili, T.; Taioe,M.; Thekisoe,O. ,and Syakalima, M.(2022). Molecular detection of zoonotic pathogens causing gastroenteritis in humans Salmonella spp., Shigella spp. and Escherichia coli isolated from Rattus species inhabiting chicken farms in North West Province, South Africa. J S Afr Vet Assoc. Online ahead of print https://doi.org/10.36303/JSAVA.83.
- Santos, R. I., Tsolis, R. M.,Baumler, A. J.,and Adams,L. G.(2003).Pathogenesis of Salmonella – induced enteritis. Brazilian Journal of Medical and Biological Research.36(1),03-12.
- Santander M.J., Roland K.L., Curtiss III R., (2008) .Regulation of Vi Capsular Polysaccharide Synthesis in Salmonella enteric Serotype Typhi J. Infect. Developing Countries, 2, 412-420.
- Song, J., Gao, X. and Galán, J. E. (2013): Structure and function of the *Salmonella Typhi chimaeric* A(2)B(5) typhoid toxin. Nature 499, 350–4.
- Kolenda ,R; Ugorski, M and Grzymajlo, K(2019). Everything You Always Wanted to Know About Salmonella Type 1 Fimbriae, but Were Afraid to Ask, Front. Microbiol. 10:1017. Doi: 10.3389/fmicb.2019.01017.
- World Health Organization (WHO). Typhoid vaccines (2018).Typhoid and other invasive salmonellosis ;(http://apps.who.in/iris/bitstream/handle/10665/2722 72/WER9313.pdf?ua=1).
- Wong ,W. ;Al Rawahi ,H. ;Patel ,S. ;Yau, Y.;Eshaghi, A.;Zittermann, S.; Tattum, L. and Morris, Sh .K.(2019).The first Canadian pediatric case of extensively drug- resistant Salmonella Typhi originating from an outbreak in Pakistan and it's the implication for empiric antimicrobial choices, Published by Elsevier I.td;https://doi.org/10.10 16/ j.idcr.2019 .e00492.
- NCBI-a

https://www.ncbi.nlm.nih.gov/protein/CHO04914.1 NCBI-b: https://www.ncbi.nlm.nih.gov/protein/UBI83717.1 NCBI-c: https://www.ncbi.nlm.nih.gov/protein/QKX99054.1

- Suardana, I. W. (2014). Analysis of Nucleotide Sequences of the 16SrRNA Gene of Novel Escherichia coli Strains Isolated from Feces of Human and Bali Cattle. J. Nucleic Acids : 1-7.
- Srinivasan, R.; Karaoz, U.; Volegova, M.; MacKichan, J.; KatoMaeda, M.; Miller, S.; Nadarajan, R.; Brodie, E. L. and Lynch, S. V. (2015). Use of 16SrRNA Gene for Identification of a Broad Range of Clinically Relevant Bacterial Pathogens . PLOS ONE. 10(2): 1-22.
- Grimont, P. A. D.; Grimont, F.; and Bouvet, P. (2000). Taxonomy of the Genus Salmonella. In: Wray, C., and Wray A. (Eds.). Salmonella in Domestic Animals. CABI Publishing, Wallingford, UK. Pp.:1-17.

- AL-Muhannak ,Fadhil Hussain Nasir(2020),Antibiotic Susceptibility and Molecular Characterization of Salmoneiia Typhi Isolated from Blood Samples in Nagaf ,Thesis ,Medical Microbiology, University of Kufa / Facuity of Medicine,15 :163.
- Zhang , Yin et al.(2018).Reciprocal Regulation of OmpR and Hfq and Their Regulatory Actions on the Vi

polysaccharide Capsular Antigen in Salmonella enterica Serovar Typhi . Current Microbiology, 75(6), pp.773-778.doi: 10.1007/s00284-018-1447-7.